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(54) ACIDES NUCLEIQUES CODANT UNE ENZYME DE PLANTE JOUANT UN ROLE DANS LA SYNTHÈSE
D'ACIDES GRAS A TRES LONGUES CHAINES

(54) NUCLEIC ACIDS ENCODING A PLANT ENZYME INVOLVED IN VERY LONG CHAIN FATTY ACID SYNTHESIS

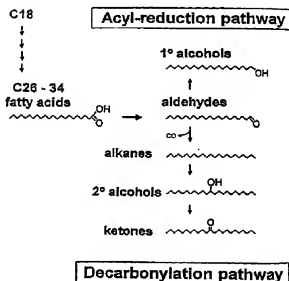
(57)

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epicuticular waxes, and methods of making such plants.



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(54) **ACIDES NUCLEIQUES CODANT UNE ENZYME DE PLANTE
JOUANT UN RÔLE DANS LA SYNTHÈSE D'ACIDES GRAS A
TRES LONGUES CHAINES**
(54) **NUCLEIC ACIDS ENCODING A PLANT ENZYME INVOLVED
IN VERY LONG CHAIN FATTY ACID SYNTHESIS**

Arabidopsis Wax Biosynthesis



(57) Molécules d'acides nucléiques codant une enzyme jouant un rôle dans l'élongation d'acides gras à très longues chaînes (VLCFA) dans des plantes. L'invention englobe un ADNc, un clone génomique et une protéine codée, ainsi que des plantes possédant une composition modifiée de VLCFA, telle que des cires épicuticulaires modifiées, et des procédés d'élaboration de ces plantes.

(57) Nucleic acid molecules encoding an enzyme involved in very long chain fatty acid (VLCFA) elongation in plants are disclosed. The invention includes a cDNA, genomic clone and encoded protein, as well as plants having modified VLCFA composition, such as modified epicuticular waxes, and methods of making such plants.



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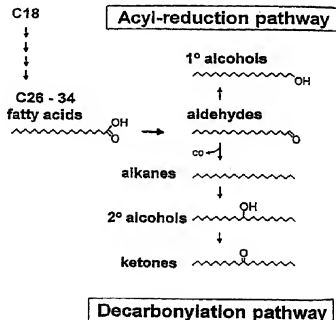
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(54) Title: NUCLEIC ACIDS ENCODING A PLANT ENZYME INVOLVED IN VERY LONG CHAIN FATTY ACID SYNTHESIS

(57) Abstract

Nucleic acid molecules encoding an enzyme involved in very long chain fatty acid (VLCFA) elongation in plants are disclosed. The invention includes a cDNA, genomic clone and encoded protein, as well as plants having modified VLCFA composition, such as modified epicuticular waxes, and methods of making such plants.

Arabidopsis Wax Biosynthesis

NUCLEIC ACIDS ENCODING A PLANT ENZYME INVOLVED IN VERY LONG CHAIN FATTY ACID SYNTHESIS

5

Technical Field

This invention relates to DNA molecules cloned from plants and methods of using such DNA molecules to produce transgenic plants with altered fatty acid composition.

Background

10 Epicuticular waxes form the outermost layer of the aerial portion of the plant and are thus the first line of interaction between the plant and its environment. The physical properties of this wax layer protect the plant from numerous environmental stresses. For example, the hydrophobic nature of wax prevents dehydration (nontomatal water loss) and aids in shedding rainwater. The reflective nature of wax protects the plant against UV radiation (Relcosky and Hanover, 1978). Waxes are also known to
15 protect against acid rain (Percy and Baker, 1990) and, because they are a good solvent for organic pollutants, they are able to impede the uptake of aqueous foliar sprays (Schreiber and Schonherr, 1992). Furthermore, surface waxes protect plants from bacterial and fungal (Jenks et al., 1994) pathogens and play a role in plant-insect interactions (Eigenbrode and Espelie, 1995). Recently it has been shown that some of the compounds found in epicuticular waxes are also present in the tryphine layer of pollen grains
20 (Preuss et al., 1993). Without these compounds the tryphine layer erodes, resulting in pollen that is unable to function causing male sterility.

 Epicuticular waxes are composed of long chain, hydrophobic compounds all derived from saturated very long chain fatty acids (VLCFAs), that are synthesized within and then secreted from the epidermis. VLCFAs are defined as those fatty acids whose chain length is 20 or more carbons long.
25 The lengths will vary from plant to plant, but typically, the wax VLCFAs are approximately 26-34 carbon long. These VLCFAs are synthesized by a microsomal fatty acid elongation (FAE) system by sequential additions of C2 moieties from malonyl-coenzyme A (CoA) to pre-existing fatty acids derived from the *de novo* fatty acid synthesis (FAS) pathway of the plastid. Analogous to *de novo* FAS it is thought that each cycle of FAE involves four enzymatic reactions; (1) condensation of malonyl-CoA with
30 a long chain acyl-CoA, (2) reduction to β -hydroxyacyl-CoA, (3) dehydration to an enoyl-CoA and (4) reduction of the enoyl-CoA, resulting in the elongated acyl-CoA (Fehling and Mukherjee, 1991). Together these four activities are termed the elongase (von Wettstein-Knowles, 1982). VLCFAs in the epidermis are then converted to the other wax components through a number of pathways consisting of multienzyme complexes. For example VLCFAs are converted to aldehydes by fatty acyl-CoA reductase
35 (Kolattukudy, 1971). These aldehydes can either be reduced by aldehyde reductase to produce primary alcohols (Kolattukudy, 1971), or decarbonylated by an aldehyde decarbonylase to produce odd chained alkanes (Cheesbrough and Kolattukudy, 1984). Alkanes can then undergo oxidation to form firstly

secondary alcohols and then ketones (for review see Post-Beittenmiller, 1996). Very little is known at the molecular level about the components that are involved in the biosynthesis of wax specific compounds and their secretion onto the plant surface. Genetic studies have shown that there are a large number of genes involved in these processes (for example, 22 loci have been reported in *Arabidopsis*, 84 in barley). However only a few of these genes have been isolated so far and the biochemical role of their gene products remains unknown (Lemieux, 1996).

In addition to being made in the epidermal cells, VLCFAs also accumulate in the seed oil of some plant species. To date, developing seeds have been the primary focus of research into VLCFA biosynthesis. In seeds VLCFAs are incorporated into triacylglycerols (TAGs), as in the *Brassicaceae*, or into wax esters, as in *Jatropha*. The seed VLCFAs include the agronomically important erucic acid (C22:1), with oils containing this fatty acid used in the manufacture of lubricants, nylon, cosmetics, pharmaceuticals and plasticisers (Battay et al., 1989; Johnston and Fritz, 1989). Conversely, VLCFAs have detrimental nutritional effects and are therefore undesirable in edible oils. This has led to the breeding of *Canola* rapeseed varieties that are almost devoid of VLCFAs (Stefansson et al., 1961).

The seeds of *Arabidopsis* contain approximately 28% [w/wt of total fatty acids (FA)] of VLCFAs, eicosenoic acid (20:1) being the predominant VLCFA (21% of wt/wt of total FA). To identify the gene products that are involved in the synthesis of seed VLCFAs and establish the VLCFA biosynthetic pathway, several groups performed mutational analysis and screened for seed that had reduced VLCFA content. Each group independently identified the *FATTY ACID ELONGATION1* gene (*FAE1*; James and Dooner, 1990; Kunst et al., 1992; Lemieux et al., 1990). A mutation at this locus resulted in reduced VLCFA levels (<1% wt/wt of total FA) in the seed. Several other mutations that were non-allelic to *FAE1* were also isolated. However, these mutations had a less pronounced effect in that VLCFAs still constituted 6.7% (wt/wt of total FA) of the seed fatty acid (Katavic et al., 1995; Kunst et al., 1992). Thus, despite the fact that four enzymatic activities are required for each elongation step, the *FAE1* gene was the only one found by mutant analysis that resulted in almost complete loss of VLCFA synthesis in the seed.

The *Arabidopsis FAE1* gene was subsequently cloned (James et al., 1995; WO 96/13582), and showed homology to three condensing enzymes: chalcone synthase, stilbene synthase and β -ketoacyl-[acyl carrier protein] synthase III (17 amino acids were identical to a 50 amino acid region of a consensus sequence for condensing enzymes). Based on this homology it was proposed that *FAE1* encodes a β -ketoacyl-coenzyme A synthase (KCS), the condensing enzyme which catalyzes the first reaction of the microsomal fatty acid elongation system (James et al., 1995). As determined by Northern analysis, the *FAE1* gene is expressed in seeds of *Arabidopsis*, but is absent from leaves (James et al., 1995). This result is consistent with the fact that the *fae1* mutation affects only the fatty acid composition of the developing seed, having no pleiotropic effects on fatty acid composition of the vegetative, or floral parts of the plant. Thus, *FAE1* is regarded as a seed-specific condensing enzyme.

Recently a cDNA from *Jatropha* seeds involved in the syntheses of VLCFAs has been isolated (Lassner et al., 1996; WO 95/15387). The protein encoded by this cDNA showed high homology to

FAEI (52% amino acid identity), and biochemical analysis demonstrated that it has a KCS activity. Using *Jojoba KCS* cDNA, Lassner et al. (1996) were able to complement the mutation in a *Canola* variety of *Brassica napus*, restoring a low erucic acid rapeseed line to a line that contained higher levels of VLCFAs. This suggests that in *Canola*, the mutation is in the structural gene encoding KCS, or a gene affecting KCS activity. Thus, both in *Arabidopsis* and *Brassica napus*, the mutations that result in the abolition of VLCFA synthesis seem to affect the condensing enzyme.

If four enzyme activities are necessary for an elongation step, and *FAEI* and *Jojoba-KCS* only encode the KCS activity, one might expect to find other complementation groups that result in very low levels of VLCFAs synthesis. Because these complementation groups were not found in mutation screenings, Millar and Kunst (1997) have hypothesized that these three activities are not seed specific, but ubiquitously present throughout the plant and shared with other FAE systems involved in VLCFA formation including wax biosynthesis. To test this *FAEI* was ecotopically expressed in yeast and in tissues of *Arabidopsis* and tobacco, where significant quantities of VLCFAs are not found. Expression of *FAEI* alone in these cells resulted in the biosynthesis and accumulation of VLCFAs. This demonstrated that the condensing enzyme is the pivotal control point of the elongase, controlling not only the amounts of VLCFAs produced, but also their chain lengths. In contrast, it appears that the other three enzyme activities of the elongase are found ubiquitously throughout the plant, are not rate limiting and play no role in the control of VLCFA synthesis. The ability of yeast containing *FAEI* to synthesize VLCFAs suggests that the expression, and the acyl chain length specificity of the condensing enzyme, along with the apparent broad specificities of the other three FAE activities, may be universal eukaryotic mechanism for regulating the amounts and acyl chain length of VLCFAs synthesized in any given cell (Millar and Kunst, 1997).

Thus, considering the central role of the condensing enzyme for VLCFA synthesis, the isolation of genes encoding condensing enzymes involved in the production of wax specific VLCFAs would facilitate the modification of wax composition through genetic engineering. Furthermore, since the majority of wax components are derived from VLCFAs, the availability of such genes would offer the potential to modify the wax load itself. This offers the potential to modify the susceptibility of plants to environmental stresses such as ultraviolet light, heat and drought, as well as the ability of plants to withstand insects and pathogens. The present invention is directed towards nucleic acids that encode condensing enzymes for VLCFA synthesis.

Summary of the Invention

The present invention provides nucleic acids (cDNAs and genomic clones) that encode a key enzyme in the synthesis of VLCFAs in plant epidermal cells. The activity of this enzyme is referred to as very long chain fatty acid elongase; the activity is required for synthesis of VLCFAs of greater than 24 carbons in length. It is shown that co-suppression of the *CUT1* gene in plants can disrupt VLCFA synthesis which results in plants having none of the protective wax usually found on stem surfaces. In addition, it is shown that such plants are conditionally male sterile: when grown under normal humidity, the plants are male sterile, but fertility can be restored by growth in an elevated humidity environment.

The invention thus provides the *CUT1* cDNA and gene nucleotide sequences ("CUT1 nucleic acids") and the amino acid sequence of the CUT1 protein. In one embodiment, the *CUT1* nucleic acids disclosed are from *Arabidopsis thaliana*. The open reading frame of the *Arabidopsis CUT1* cDNA molecule encodes an enzyme of 497 amino acids which catalyzes the addition of 2C units to pre-existing C24 or longer fatty acids.

Also encompassed within the scope of this invention are transformation vectors that include at least a portion of the *CUT1* nucleic acid molecules. Such vectors may be transformed into plants to produce transgenic plants with modified VLCFA compositions (relative to non-transgenic plants of the same species). Depending on the particular sequences incorporated into the vector, transformation with the *CUT1* cDNA, gene or derivatives thereof can be used to modify agronomically important traits, including the presence, composition and thickness of epicuticular wax layers on leaves and stems, seed coat fatty acids, seed oil composition and male sterility. Typically, such vectors include regulatory sequences, such as promoters, operably linked to the *CUT1* open reading frame or a derivative of the *CUT1* nucleic acids. For example, VLCFA synthesis may be altered by introducing into a plant a transformation vector that includes a sense or antisense version of the *CUT1* cDNA. Transgenic plants having modified VLCFA compositions and which are transformed with such recombinant transformation vectors are also provided by this invention.

In one aspect of the invention, transformation with sense or antisense versions of the *CUT1* nucleic acids may be used to produce plants having modified epicuticular wax layers on the aerial parts of the plants, such as the leaves and stems. A modified epicuticular wax layer may be modified in physical respects, such as thickness of the wax layer, or in composition. Because these layers play a role in the ability of plants to resist environmental stresses, such as drought and ultraviolet light, as well as insects and pathogens, transformation with vectors including forms of the *CUT1* nucleic acids may be used to produce plants with particular agronomic advantages. Producing plants with modified epicuticular wax composition may be achieved by introducing into the plants a vector in which the *CUT1* nucleic acid (or a derivative thereof) is operably linked to a promoter that directs expression of the open reading frame in the epidermal cells. The CaMV 35S promoter and the endogenous *CUT1* gene promoter are examples of regulatory sequences that may be suitable for this purpose.

Agronomically important traits in addition to wax composition may also be modified using the *CUT1* nucleic acids of the present invention. For example, the fatty acid composition of the seed coat and the fatty acid composition of seed oil may be modified by transforming plants with the *CUT1* cDNA or derivatives thereof. Preferably, where it is desired to modify aspects of seed VLCFA composition, the introduced *CUT1* nucleic acid sequence will be operably linked to a promoter known to direct expression in seed tissues. Seed-specific promoters include the napin promoter of *Brassica napus* (Lee et al., 1991). In addition, transformation with the *CUT1* nucleic acids or derivatives thereof may be used to disrupt VLCFA synthesis in pollen, resulting in conditionally male sterile plants. Such plants are useful in plant breeding programs.

While the invention provides *CUT1*-encoding nucleic acids from *Arabidopsis*, it additionally

encompasses homologs, orthologs and variants and derivatives of these sequences, as well as homologs, orthologs and variants of the CUT1 polypeptide sequence. Thus, in one aspect of the invention, nucleic acid molecules that comprise specified regions of these sequences are provided. Exemplary of such nucleic acid molecules are oligonucleotides that are useful as probes or primers to detect and amplify *CUT1*-
 5 encoding nucleic acids from other plant species. Such oligonucleotides are useful as hybridization probes or PCR primers, and typically comprise at least 15 consecutive bases of the disclosed *CUT1* nucleic acid sequences. In other embodiments, such oligonucleotides comprise longer regions of the disclosed *CUT1* sequences, such as at least 20, 25 or 30 consecutive nucleotides.

In another aspect, the invention provides compositions and methods for isolating nucleic acid
 10 sequences that encode enzymes having CUT1 activity from other plant species. Typically, such methods involve hybridizing probes or primers derived from the disclosed *Arabidopsis* sequences to nucleic acids obtained or derived from such other plant species.

Homologous and orthologous sequences to *Arabidopsis CUT1* nucleic acid and CUT1 amino acid sequences share key functional and structural characteristics with the disclosed *Arabidopsis* sequences.
 15 Functionally, such sequences encode (or comprise) a polypeptide that catalyzes the very long chain fatty acid elongation as described above. Structurally, such sequences share a specified structural relationship with the disclosed sequences. By way of example, in certain embodiments, homologous amino acid sequences have at least 70% sequence identity with the *Arabidopsis CUT1* amino acid sequence. In other embodiments, homologous nucleic acid sequences hybridize under stringent conditions to the disclosed
 20 *Arabidopsis CUT1* nucleic acid sequences.

Another aspect of the invention relates to the purified CUT1 enzyme itself. Having provided nucleic acid molecules that encode this enzyme, the invention also facilitates the expression of CUT1 enzyme in heterologous systems, including *E. coli*, yeast and baculovirus expression systems. Thus, the invention permits the large scale production of the enzyme for agricultural and other applications.

25 In another aspect of the invention the promoter sequence of the *CUT1* gene is disclosed. This promoter sequence confers epidermis-specific expression, and may be used to express a variety of nucleic acids in an epidermis-specific manner.

Detailed Description of the Invention

30 I. Definitions

Unless otherwise noted, technical terms are used according to conventional usage. Definitions of common terms in molecular biology may be found in Benjamin Lewin, *Genes V* published by Oxford University Press, 1994 (ISBN 0-19-854287-9); Kendrew et al (eds.), *The Encyclopedia of Molecular Biology*, published by Blackwell Science Ltd., 1994 (ISBN 0-632-02182-9); and Robert A. Meyers (ed.),
 35 *Molecular Biology and Biotechnology: a Comprehensive Desk Reference*, published by VCH Publishers, Inc., 1995 (ISBN 1-56081-569-8). The nomenclature for DNA bases as set forth at 37 CFR § 1.822 and the standard three letter codes for amino acid residues are used herein.

In order to facilitate review of the various embodiments of the invention, the following

definitions of terms are provided:

CUT1 protein: The defining functional characteristic of a CUT1 protein is its enzymatic activity, specifically its very long chain fatty acid elongase activity. This activity is manifested as the catalysis of one or more steps in the addition of 2 carbon moieties (such as malonyl-coenzyme A) to pre-existing very long chain fatty acids (VLCFAs). In a preferred embodiment, a CUT1 protein catalyzes one or more steps in the addition of 2 carbon moieties to pre-existing long chain fatty acids of at least 24 carbon units in length. This activity can be measured by the assay described below.

This invention provides a cDNA and a gene encoding a CUT1 enzyme from *Arabidopsis thaliana*. However the invention is not limited to this particular CUT1 protein: other nucleotide sequences which encode CUT1 proteins are also part of the invention, including variants on the disclosed *Arabidopsis* cDNA and gene sequences and orthologous sequences from other plant species, including naturally occurring variants, such as sequences from other ecotypes, species and natural polymorphisms, the cloning of which is now enabled. Such sequences share the essential functional characteristic of encoding an enzyme having very long chain fatty acid elongase activity. Nucleic acid sequences that encode CUT1 proteins and the proteins encoded by such nucleic acids share not only this functional characteristic, but also a specified level of sequence similarity (or sequence identity), as addressed below. The concept of sequence identity can also be expressed in the ability of two sequences to hybridize to each other under stringent conditions.

Sequence identity: the similarity between two nucleic acid sequences, or two amino acid sequences is expressed in terms of the similarity between the sequences, otherwise referred to as sequence identity. Sequence identity is frequently measured in terms of percentage identity (or similarity or homology); the higher the percentage, the more similar the two sequences are.

Methods of alignment of sequences for comparison are well-known in the art. Various programs and alignment algorithms are described in: Smith and Waterman (1981); Needleman and Wunsch (1970); Pearson and Lipman (1988); Higgins and Sharp (1988); Higgins and Sharp (1989); Corpet et al. (1988); and Pearson et al. (1994). Altschul et al. (1994) presents a detailed consideration of sequence alignment methods and homology calculations.

The NCBI Basic Local Alignment Search Tool (BLAST) (Altschul et al., 1990) is available from several sources, including the National Center for Biological Information (NCBI, Bethesda, MD) and on the Internet, for use in connection with the sequence analysis programs blastp, blastn, blastx, tblastn and tblastx. It can be accessed at <http://www.ncbi.nlm.nih.gov/BLAST/>. A description of how to determine sequence identity using this program is available at http://www.ncbi.nlm.nih.gov/BLAST/blast_help.html.

Homologs of the *Arabidopsis* CUT1 protein are characterized by possession of at least 70% sequence identity counted over the full length alignment with the disclosed *Arabidopsis* CUT1 amino acid sequence using the NCBI Blast 2.0, gapped blastp set to default parameters. Such homologous peptides will more preferably possess at least 75%, more preferably at least 80% and still more preferably at least 90% or 95% sequence identity with the *Arabidopsis* CUT1 amino acid sequence determined by this method. When less than the entire sequence is being compared for sequence identity, homologs will

possess at least 75% and more preferably at least 85% and more preferably still at least 90% or 95% sequence identity over short windows of 10-20 amino acids. Methods for determining sequence identity over such short windows are described at http://www.ncbi.nlm.nih.gov/BLAST/blast_FAQs.html.

- Homologs having the sequence identities described above will, in some embodiments, also possess
- 5 VLCFA elongase activity. One of skill in the art will appreciate that these sequence identity ranges are provided for guidance only; it is entirely possible that strongly significant homologs could be obtained that fall outside of the ranges provided. The present invention provides not only the peptide homologs are described above, but also nucleic acid molecules that encode such homologs.

- Homologs of the *Arabidopsis CUT1* cDNA and gene are similarly characterized by possession of
- 10 at least 60% sequence identity counted over the full length alignment with the disclosed *Arabidopsis* cDNA or gene sequence using the NCBI Blast 2.0, gapped blastn set to default parameters. Such homologous nucleic acids will more preferably possess at least 70%, more preferably at least 80% and still more preferably at least 90% or 95% sequence identity determined by this method. When less than the entire sequence is being compared for sequence identity, homologs will possess at least 85% and more
- 15 preferably at least 90% and more preferably still at least 95% sequence identity over 30 nucleotide windows. Homologs having the sequence identities described above will, in some embodiments, also encode a polypeptide having VLCFA elongase activity. However, homologs as defined above are useful for modifying VLCFA elongase activity in transgenic plants (for example, as used in antisense constructs) even when they do not encode a functional peptide. Again, one of skill in the art will appreciate that these
- 20 sequence identity ranges are provided for guidance only; it is entirely possible that strongly significant nucleic acid homologs could be obtained that fall outside of the ranges provided.

- Another indication that two nucleic acid molecules are substantially homologous is that the two molecules hybridize to each other under stringent conditions when one molecule is used as a hybridization probe, and the other is present in a biological sample, e.g., genomic material from a cell. Specific
- 25 hybridization means that the molecules hybridize substantially only to each other and not to other molecules that may be present in the genomic material. Stringent conditions are sequence dependent and are different under different environmental parameters. Generally, stringent conditions are selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the
- 30 target sequence hybridizes to a perfectly matched probe. Conditions for nucleic acid hybridization and calculation of stringencies can be found in Sambrook et al. (1989) and Tijssen (1993). Hybridization conditions and stringencies are further discussed below.

- Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences, due to the degeneracy of the genetic code. It is understood that changes in
- 35 nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequence that all encode substantially the same protein.

Probes and primers: Nucleic acid probes and primers may readily be prepared based on the nucleic acids provided by this invention. A probe comprises an isolated nucleic acid attached to a

detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. Methods for labeling and guidance in the choice of labels appropriate for various purposes are discussed, e.g., in Sambrook et al. (1989) and Ausubel et al. (1987).

Primers are short nucleic acids, preferably DNA oligonucleotides 15 nucleotides or more in length. Primers may be annealed to a complementary target DNA strand by nucleic acid hybridization to form a hybrid between the primer and the target DNA strand, and then extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR) or other nucleic-acid amplification methods known in the art.

Methods for preparing and using probes and primers are described, for example, in Sambrook et al. (1989), Ausubel et al. (1987), and Innis et al., (1990). PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, © 1991, Whitehead Institute for Biomedical Research, Cambridge, MA). One of skill in the art will appreciate that the specificity of a particular probe or primer increases with its length. Thus, for example, a primer comprising 20 consecutive nucleotides of the *Arabidopsis CUT1* cDNA or gene will anneal to a target sequence (e.g., a corresponding *CUT1* gene from *Zea mays*) with a higher specificity than a corresponding primer of only 15 nucleotides. Thus, in order to obtain greater specificity, probes and primers may be selected that comprise 20, 25, 30, 35, 40, 50 or more consecutive nucleotides of the *Arabidopsis CUT1* cDNA or gene sequences. Such probes and primers are useful for obtaining *CUT1* nucleic acid molecules (cDNA, genomic sequences, and portions of these molecules) both from *Arabidopsis* and other plant species.

Vector: A nucleic acid molecule as introduced into a host cell, thereby producing a transformed host cell. A vector may include nucleic acid sequences that permit it to replicate in the host cell, such as an origin of replication. A vector may also include one or more selectable marker genes and other genetic elements known in the art.

Transformed: A transformed cell is a cell into which has been introduced a nucleic acid molecule by molecular biology techniques. As used herein, the term transformation encompasses all techniques by which a nucleic acid molecule might be introduced into such a cell, including transformation with *Agrobacterium* vectors, transfection with viral vectors, transformation with plasmid vectors, and introduction of naked DNA by electroporation, lipofection, and particle gun acceleration.

Isolated: An "isolated" biological component (such as a nucleic acid or protein) has been substantially separated or purified away from other biological components in the cell of the organism in which the component naturally occurs, i.e., other chromosomal and extrachromosomal DNA and RNA, and proteins. Nucleic acids and proteins which have been "isolated" thus include nucleic acids and proteins purified by standard purification methods. The term also embraces nucleic acids and proteins prepared by recombinant expression in a host cell as well as chemically synthesized nucleic acids.

Purified: The term purified does not require absolute purity; rather, it is intended as a relative term. Thus, for example, a purified *CUT1* protein preparation is one in which the *CUT1* protein is more

enriched than the protein is in its natural environment within a cell. Preferably, a preparation of CUT1 protein is purified such that CUT1 protein represents at least 50% of the total protein content of the preparation.

Operably linked: A first nucleic acid sequence is operably linked with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter effects the transcription or expression of the coding sequence. Generally, operably linked DNA sequences are contiguous and, where necessary to join two protein coding regions, in the same reading frame.

Recombinant: A recombinant nucleic acid is one that has a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques.

Ortholog: two nucleotide or amino acid sequences are orthologs of each other if they share a common ancestral sequence and diverged when a species carrying that ancestral sequence split into two species. Orthologous sequences are also homologous sequences.

Transgenic plant: as used herein, this term refers to a plant that contains recombinant genetic material not normally found in plants of this type and which has been introduced into the plant in question (or into progenitors of the plant) by human manipulation. Thus, a plant that is grown from a plant cell into which recombinant DNA is introduced by transformation is a transgenic plant, as are all offspring of that plant which contain the introduced DNA (whether produced sexually or asexually).

II. Sequence Listing and Figures

The nucleic and amino acid sequences listed in the accompanying sequence listing are showed using standard letter abbreviations for nucleotide bases, and three letter code for amino acids. Only one strand of each nucleic acid sequence is shown, but the complementary strand is understood to be included by any reference to the displayed strand.

Seq. I.D. No. 1 shows the nucleotide sequence of the *CUT1* gene and the encoded amino acid sequence.

Seq. I.D. No. 2 shows the nucleotide sequence of the *CUT1* cDNA.

Seq. I.D. No. 3 shows the nucleotide sequence of the *CUT1* open reading frame.

Seq. I.D. No. 4 shows the amino acid sequence of the CUT1 protein.

Seq. I.D. Nos. 5 - 11 show primers useful in PCR amplification of various regions of the *CUT1* gene, cDNA or ORF.

Seq. I.D. No.12 shows the promoter region of the *CUT1* genomic clone.

Fig. 1 shows the pathways of wax biosynthesis in *Arabidopsis*.

III. Isolation and Characterization of the *CUT1* cDNA

5 The *CUT1* cDNA was initially identified using a TBLASTN homology search (Altschul et al., 1990) of the database of expressed sequenced tags (ESTs) of anonymous *Arabidopsis* cDNA clones (Newman et al., 1994) using the deduced amino acid sequence of the *FAE1* gene. The search found 14 ESTs in the database which had open reading frames with significant homology to *FAE1*. These ESTs did not correspond to known condensing enzymes such as chalcone synthase or 3-ketoacyl-acyl carrier
10 protein synthase III.

One of these ESTs was selected for further investigation, and the corresponding full length cDNA was isolated. This cDNA is herein referred to as the *CUT1* cDNA. Sequencing demonstrated that the *CUT1* cDNA was 1829 nucleotides long, approximately the size of the *FAE1* transcript (James et al., 1995). The *CUT1* cDNA contains one open reading frame of 497 amino acids, which is shorter than
15 both the *FAE1* sequence (506 amino acids) and the jojoba KCS (521 amino acids). The *CUT1* cDNA and the protein it encodes are shown in Seq. I.D. Nos. 2 and 4, respectively.

There is an in frame stop codon, TAA, 15 nucleotides upstream of the most 5' ATG, suggesting that this sequence indeed represents the full length amino acid sequence of the protein. Thus, the *CUT1* cDNA as depicted in Seq. I.D. No. 2 has a 5' untranslated region of 58 nucleotides, an open reading
20 frame of 1491 nucleotides and a 3' untranslated region of 258 nucleotides, excluding the poly(A) tail (22 As). Comparison of the deduced amino acid sequence of the *CUT1* protein to *FAE1* revealed that they are 50.0% identical and 74.7% similar.

IV. Isolation and Characterization of the *CUT1* Gene

25 An *Arabidopsis CUT1* genomic clone was isolated from a genomic library in λ GEM11 by probing nitrocellulose plaque lifts with a full-length *CUT1* cDNA clone. A 2.5 kb long *SalI* fragment containing 580 bp of the coding sequence and 1951 bp of the 5' upstream region was subcloned into the *SalI* site of pT773 18U plasmid (Pharmacia), followed by complete sequencing on both strands. The sequence of this genomic clone is shown in Seq. I.D. No. 1.

30 *In situ* hybridization studies in developing shoots, leaves and siliques of *Arabidopsis* indicated epidermis-specific expression of the *CUT1* gene, as expected of a gene encoding an enzyme involved in wax biosynthesis.

V. Analysis of the *CUT1* Promoter

35 In order to confirm the tissue and cell specificity of the *CUT1* promoter, 5' flanking sequences from the *CUT1* genomic clone were operably linked to the *uidA* reporter gene encoding β -glucuronidase (GUS). Two constructs were made, one having a 1.9 kb promoter fragment and the second containing a truncated 1.2 kb promoter. These promoter-GUS fusions were introduced into *Arabidopsis* and tobacco by *Agrobacterium*-mediated transformation and the promoter function characterized in transgenic plants.

40 To obtain the 1.9 and 1.2 kb regions of the *CUT1* promoter sequence, synthetic oligonucleotides

homologous to portions of the 5' untranslated region of the genomic clone were used as primers to amplify either a 1949 bp or a 1209 bp promoter fragment by PCR. As shown in Figure 1, the upstream primer was 5'-GTGCTTTATATATGTTTG-3' (cutpro3) (Seq. I.D. No. 5) in combination with the downstream primer 5'-CGTGGGAGAGTTTAAATG-3' (cutpro1) (Seq. I.D. No. 6) for the PCR-synthesis of the 1949 bp fragment, and 5'-CTTCGATATCGGTTGTTG-3' (cutpro2) (Seq. I.D. No. 7) and cutpro1 for the amplification of the 1209 bp fragment. In both cases, the amplified products were subcloned in the *HincII* site of the plasmid pT7T318U (Pharmacia). The inserts were then cleaved out with *HindIII* and *XbaI* and directionally subcloned into the corresponding sites of the binary Ti plasmid pBI101 (Clontech), which contains a promoterless GUS gene (Jefferson et al. 1987). The *pCUT1*-GUS fusion constructs in pBI101 were introduced into *Agrobacterium tumefaciens* strain GV3101 (Koncz and Scheil, 1986) by electroporation and selected for resistance to kanamycin (50 µg/ml).

For transformation of tobacco, *Agrobacterium* harbouring the *pCUT1*-GUS construct was co-cultivated with leaf pieces of *Nicotiana tabacum* SR1 and transformants were selected with kanamycin (100mg/mL) on solid medium (Lee and Douglas, 1996). *Arabidopsis thaliana* (L.) Heynh. ecotype Columbia was transformed with *pCUT1*-GUS binary vector using a combination of *in planta* (Chang et al., 1994, Katavic et al., 1994) and vacuum infiltration methods (Bechtold et al., 1993). Plants were grown until the primary inflorescence shoots reached 1-2 cm in height, when this bolts were cut off. The wound site was inoculated with 50 mL of an overnight *Agrobacterium* culture. After 4-6 days a number of secondary inflorescences that appeared were cut off, and vacuum infiltration was performed on these plants using the conditions described by Bechtold et al. (1993). Screening for transformed seed was done on 50µg/mL kanamycin as described previously (Katavic et al., 1994).

Tissue sections of transgenic plants containing the *pCUT1*-GUS constructs were placed in 100 mM NaPO₄ (pH7) and 1 mM spermidine for 15 min, then incubated at 37° C in 0.5 K₂[Fe(CN)₆], 0.01 % Triton X-100, 1mM EDTA, 10 mM β-mercaptoethanol, 5-bromo-4-chloro-3-indolyl-β-D-glucuronide in 100 mM NaPO₄ (pH7), until a blue color appeared (after approximately 1 hr). Following incubation with the substrate, chlorophyll was removed from the sections using a graded ethanol series.

In both recipient plant species, *Arabidopsis* and tobacco, *CUT1* expression pattern mirrored that observed in the *in situ* experiments. Furthermore, both long and short *CUT1* promoter fragments targeted expression of the *uidA* gene exclusively to the epidermis. No GUS expression was detected in any of the other cell types in the stems or leaves of transgenic plants. Thus, the *Arabidopsis CUT1* promoter is regulated in a tissue specific, and cell specific manner, and epidermis specificity appears to be retained even in unrelated plant species like tobacco. In addition, no differences in the strength of expression were detected between the 1.9 kb and 1.2 kb promoter.

35 VI. Preferred Methods for Producing *CUT1* Nucleic Acids

With the provision of the *CUT1* cDNA and gene (the "*CUT1* nucleic acids") the polymerase chain reaction (PCR) may now be utilized in a preferred method for producing the *CUT1* nucleic acids. PCR amplification of the *CUT1* cDNA sequence may be accomplished either by direct PCR from a plant

cDNA library or by Reverse-Transcription PCR (RT-PCR) using RNA extracted from plant cells as a template. Methods and conditions for both direct PCR and RT-PCR are known in the art and are described in Innis et al. (1990). Suitable plant cDNA libraries for direct PCR include the *Arabidopsis* cDNA library described by Newman et al. (1994). Similarly, the *CUT1* genomic sequence may be amplified directly from genomic DNA extracted from plants, or from plant genomic DNA libraries. Amplification may be used to obtain the full length cDNA or genomic sequence, or may be used to amplify selected portions of these molecules (for example for use in antisense constructs)

The selection of PCR primers will be made according to the portions of the *CUT1* nucleic acids which are to be amplified. Variations in amplification conditions may be required to accommodate primers of differing lengths; such considerations are well known in the art and are discussed in Innis et al. (1990), Sambrook et al. (1989), and Ausubel et al. (1987). By way of example only, the entire *CUT1* cDNA molecule as shown in Seq. I.D. No. 2 may be amplified using the following combination of primers:

primer 1 5' AAATACCCTAATCACATTTGTAA 3' (Seq. I.D. No. 8)

primer 2 5' TTAAACAGAGAGAAATATTCCTA 3' (Seq. I.D. No. 9)

The open reading frame portion of the cDNA may be amplified using the following primer pair:

primer 3 5' ATGCCTCAGGCACCGATGCCAGAG 3' (Seq. I.D. No. 10)

primer 4 5' CAGCAGGAGAACTAAAAAATACC 3' (Seq. I.D. No. 11)

These primers are illustrative only; it will be appreciated by one skilled in the art that many different primers may be derived from the provided sequences in order to amplify particular regions of the *CUT1* sequences. Resequencing of PCR products obtained by these amplification procedures is recommended; this will facilitate confirmation of the amplified *CUT1* sequence and will also provide information on natural variation on this sequence in different ecotypes and plant populations.

Oligonucleotides which are derived from the *CUT1* nucleic acid sequences and which are suitable for use as PCR primers to amplify the *CUT1* nucleic acid sequences are encompassed within the scope of the present invention. Preferably, such oligonucleotide primers will comprise a sequence of 15-20 consecutive nucleotides of the *CUT1* nucleic acid sequences. To enhance amplification specificity, primers comprising at least 20-30 consecutive nucleotides of these sequences may also be used.

VII. Cloning *CUT1* Variants

With the provision herein of the *CUT1* nucleic acid sequences, the cloning by standard methodologies of corresponding cDNAs and genes from other ecotypes and plant species, as well as polymorphic forms of the disclosed sequences is now enabled. Thus, the present invention includes methods of isolating a nucleotide sequence encoding a plant very long chain fatty acid elongation enzyme from a plant. Both conventional hybridization and PCR amplification procedures may be utilized to clone such sequences. Common to both of these techniques is the hybridization of probes or primers derived from the disclosed *CUT1* nucleic acid sequences to a target nucleotide preparation, which may

be, in the case of conventional hybridization approaches, a cDNA or genomic library or, in the in the case of PCR amplification, extracted genomic DNA, mRNA, a cDNA library or a genomic library.

Direct PCR amplification may be performed on cDNA libraries prepared from the plant species in question, or RT-PCR may be performed using mRNA extracted from the plant cells using standard methods. PCR primers will comprise at least 15 consecutive nucleotides of the *CUT1* nucleic acid sequences. One of skill in the art will appreciate that sequence differences between the disclosed *CUT1* nucleic acid sequences and the target gene to be amplified may result in lower amplification efficiencies. To compensate for this, longer PCR primers or lower annealing temperatures may be used during the amplification cycle. Where lower annealing temperatures are used, sequential rounds of amplification using nested primer pairs may be necessary to enhance specificity.

For conventional hybridization techniques, the hybridization probe is preferably labeled with a detectable label such as a radioactive label, and the probe is of at least 20 nucleotides in length. As is well known in the art, increasing length of hybridization probes tends to give enhanced specificity. The labeled probe derived from, for example, the *CUT1* cDNA sequence may be hybridized to a plant cDNA or genomic library and the hybridization signal detected using means known in the art. The hybridizing colony or plaque (depending on the type of library used) is then purified and the cloned sequence contained in that colony or plaque isolated and characterized.

VIII. Use of the *CUT1* Nucleic Acids to Produce Plants with Modified VLCFA Composition

Once a gene or cDNA ("nucleic acid") encoding a protein involved in the determination of a particular plant characteristic has been isolated, standard techniques may be used to express the nucleic acid in transgenic plants in order to modify that particular plant characteristic. The basic approach is to clone the nucleic acid into a transformation vector, such that it is operably linked to control sequences (e.g., a promoter) which direct expression of the open reading frame in plant cells. The transformation vector is then introduced into plant cells by one of a number of techniques (e.g., electroporation) and progeny plants containing the introduced nucleic acid are selected. Preferably all or part of the transformation vector will stably integrate into the genome of the plant cell. That part of the transformation vector which integrates into the plant cell and which contains the introduced nucleic acid and associated sequences for controlling expression (the introduced "transgene") may be referred to as the recombinant expression cassette.

Selection of progeny plants containing the introduced transgene may be made based upon the detection of an altered phenotype. Such a phenotype may result directly from the nucleic acid cloned into the transformation vector or may be manifested as enhanced resistance to a chemical agent (such as an antibiotic) as a result of the inclusion of a dominant selectable marker gene incorporated into the transformation vector.

The choice of (a) control sequences and (b) how the nucleic acid (or selected portions of the nucleic acid) are arranged in the transformation vector relative to the control sequences determine, in

part, how the plant characteristic affected by the introduced nucleic acid is modified. For example, the control sequences may be tissue specific, such that the nucleic acid is only expressed in particular tissues of the plant (e.g., pollen) and so the affected characteristic will be modified only in those tissues. The nucleic acid sequence may be arranged relative to the control sequence such that the nucleic acid transcript is expressed normally, or in an antisense orientation. Expression of an antisense RNA corresponding to the cloned nucleic acid will result in a reduction of the targeted gene product (the targeted gene product being the protein encoded by the plant gene from which the introduced nucleic acid was derived). Over-expression of the introduced nucleic acid, resulting from a plus-sense orientation of the nucleic acid relative to the control sequences in the vector, may lead to an increase in the level of the gene product, or may result in co-suppression (also termed "sense suppression") of that gene product.

Successful examples of the modification of plant characteristics by transformation with cloned nucleic acid sequences are replete in the technical and scientific literature. Selected examples, which serve to illustrate the current knowledge in this field of technology, and which are herein incorporated by reference, include:

U.S. Patent No. 5,451,514 to Boudet (modification of lignin synthesis using antisense RNA and co-suppression);

U.S. Patent No. 5,443,974 to Hitz (modification of saturated and unsaturated fatty acid levels using antisense RNA and co-suppression);

U.S. Patent No. 5,530,192 to Murase (modification of amino acid and fatty acid composition using antisense RNA);

U.S. Patent No. 5,455,167 to Voelker (modification of medium chain fatty acids)

U.S. Patent No. 5,231,020 to Jorgensen (modification of flavonoids using co-suppression);

U.S. Patent No. 5,583,021 to Dougherty (modification of virus resistance by expression of plus-sense untranslatable RNA);

WO 96/13582 (modification of seed VLCFA composition using over expression, co-suppression and antisense RNA in conjunction with the *Arabidopsis* FAE1 gene); and

WO 95/15387 (modification of seed VLCFA composition using over expression of jojoba wax synthesis gene).

These examples include descriptions of transformation vector selection, transformation techniques and the construction of constructs designed to over-express the introduced nucleic acid or to express antisense RNA corresponding to the nucleic acid. In light of the foregoing and the provision herein of the *CUT1* nucleic acids, it is thus apparent that one of skill in the art will be able to introduce these nucleic acids, or derivative forms of these molecules (e.g., antisense forms), into plants in order to produce plants having modified VLCFA compositions. Examples one and two below provides illustrations of this in which the *CUT1* cDNA is operably linked to the CaMV 35S promoter sequence, cloned into the pBIN19 transformation vector and introduced into *Arabidopsis* using a vacuum infiltration method.

As reported in Example one, certain of the plants transformed in this way had no detectable epicuticular wax layers, indicating that transformation with the *CUT1* cDNA had disrupted normal VLCFA synthesis in the plant epidermal cells. Such disruption is likely attributable to the phenomenon termed co-suppression (or sense-suppression). These plants are thus referred to as "*CUT1*-suppressed".

- 5 This phenomenon may be affected by factors such as positional location of the introduced sequences in the plant genome.

Over-expression of CUT1 protein in transgenic plants, resulting in plants enhanced epicuticular wax layers will be a useful agronomic trait, providing increased drought and insect resistance. For example, drought resistance in rice is associated with high wax lines rich in C₂₉, C₃₁ and C₃₃ alkanes
 10 (O'Toole and Cruz, 1983; Haque et al., 1992). Increased wax deposition in transgenic plants can be accomplished by overexpression of CUT1 protein, while the identification of the *CUT1* promoter allows targeting of lipid modification enzymes such as desaturases, thioesterases and other condensing enzymes with different specificities to the epidermal cells to modify wax composition.

Transformation of plants with the *CUT1* nucleic acids or derivatives thereof may be used to
 15 modify other plant characteristics, such as seed coat composition and seed oil composition. Because condensing enzymes are pivotal enzymes in the synthesis of VLCFAs, controlling levels of accumulation of VLCFAs and their acyl chain length (Millar and Kunst, 1997) through the manipulation of CUT1 expression will permit the production of plants having novel fatty acid compositions. For instance, the accumulation of VLCFAs in tobacco seed expressing *FAE1* from *Arabidopsis* (Millar and Kunst, 1997)
 20 raises the possibility of producing VLCFAs in plant species that currently do not synthesize VLCFAs. In addition, targeting of CUT1 to seeds will be useful to produce crop plants capable of synthesizing new, agronomically important VLCFAs in seed oil.

Disruption of CUT1 activity in transgenic plants also provides a simple means for obtaining conditional male sterility in plants (see Example two). One of the major factors contributing to increases
 25 in crop productivity is the development of hybrid varieties of crops. Several different breeding strategies have been used to produce hybrid seed, but none of these strategies can be used as a general approach in all crop plants (Goldberg et al., 1993). As an alternative, genetically engineered systems and strategies for male fertility control that are applicable to a wide range of crops have recently been developed. For example, nuclear male sterility has been engineered by (1) tapetum-specific expression of a bacterial
 30 RNase gene (Mariani et al., 1990, 1992), (2) overexpression of the *rolC* gene from *Agrobacterium rhizogenes* (Fladung, 1990; Schmülling et al., 1988, 1992), (3) expression of glucanase that disrupts the callose wall of the microsporophyte prematurely (Tsuchiya et al., 1995; Worrall et al., 1992), (4) the inhibition of flavonoid biosynthetic genes like chalcone synthase and dihydroflavonol 4-reductase (van der Krol et al., 1988, 1990; van der Meer et al., 1992; Napoli et al. 1990; Taylor and Jorgensen, 1992), and (5)
 35 altered expression of stilbene synthase (Fischer et al., 1997). However, in most of these cases the restoration of fertility is not simple, or not easily controlled. In contrast, conditional male sterility caused by suppression of CUT1 activity is easily reversible under high relative humidity.

The selection of vectors and promoters appropriate for targeting particular characteristics for

modification (such as seed-specific expression) are well known; the following paragraphs set forth general guidance on the various options available in producing transgenic plants having modified VLCFA composition.

5 a. Plant Types

VLCFAs are found in all plant types, and thus DNA molecules according to the present invention (e.g., the *CUT1* cDNA, gene, homologs and antisense forms thereof) may be introduced into any plant type in order to modify the VLCFA composition of the plant. Thus, the sequences of the present invention may be used to modify VLCFA composition in any higher plant, including
10 monocotyledonous and dicotyledonous plants, including, but not limited to maize, wheat, rice, barley, soybean, beans in general, rape/canola, alfalfa, flax, sunflower, safflower, brassica, cotton, flax, peanut, clover; vegetables such as lettuce, tomato, cucurbits, potato, carrot, radish, pea, lentils, cabbage, broccoli, brussel sprouts, peppers; tree fruits such as apples, pears, peaches, apricots; flowers such as carnations and roses.

15

 b. Vector Construction, Choice of Promoters

A number of recombinant vectors suitable for stable transfection of plant cells or for the establishment of transgenic plants have been described including those described in Pouwels et al., (1987), Weissbach and Weissbach, (1989), and Gelvin et al., (1990). Typically, plant transformation
20 vectors include one or more cloned plant genes (or cDNAs) under the transcriptional control of 5' and 3' regulatory sequences and a dominant selectable marker. Such plant transformation vectors typically also contain a promoter regulatory region (e.g., a regulatory region controlling inducible or constitutive, environmentally-or developmentally-regulated, or cell- or tissue-specific expression), a transcription initiation start site, a ribosome binding site, an RNA processing signal, a transcription termination site,
25 and/or a polyadenylation signal.

Examples of constitutive plant promoters which may be useful for expressing *CUT1* nucleic acids include: the cauliflower mosaic virus (CaMV) 35S promoter, which confers constitutive, high-level expression in most plant tissues (see, e.g., Odel et al., 1985, Dekeyser et al., 1990, Terada and Shimamoto, 1990); the nopaline synthase promoter (An et al., 1988); and the octopine synthase promoter
30 (Fromm et al., 1989).

A variety of plant gene promoters that are regulated in response to environmental, hormonal, chemical, and/or developmental signals, also can be used for expression of *CUT1* nucleic acids in plant cells, including promoters regulated by: (a) heat (Callis et al., 1988); (b) light (e.g., the pea *rbcs-3A* promoter, Kuhlmeier et al., 1989, the maize *rbcs* promoter, Schaffner and Sheen, 1991, and the
35 chlorophyll *a/b* binding protein promoter, Simpson et al., 1985); (c) hormones, such as abscisic acid (Marcotte et al., 1989); (d) wounding (e.g., *wund*, Siebertz et al., 1989); and (e) chemicals such as methyl jasmonate or salicylic acid. It may also be advantageous to employ tissue-specific promoters, such as those described by Roshal et al., (1987), Scherthaner et al., (1988), and Bustos et al., (1989).

Alternatively, tissue specific (root, leaf, flower, and seed for example) promoters (Carpenter et al. 1992, Denis et al. 1993, Opperman et al. 1993, Stockhouse et al. 1997; Roshal et al., 1987; Scherthner et al., 1988; and Bustos et al., 1989) can be fused to the coding sequence to obtain particular expression in respective organs. In addition, the timing of the expression can be controlled by using promoters such as those acting at senescencing (Gan and Amasino 1995) or late seed development (Odell et al. 1994). The promoter region of the *CUT1* genomic sequence disclosed herein confers epidermis-specific expression in *Arabidopsis* and tobacco. Accordingly, the native promoter may be used to obtain epidermis-specific expression of the introduced transgene.

For producing conditionally male sterile plants by blocking *CUT1* activity in pollen, it is preferable to use a pollen-specific promoter (so as to avoid pleiotropic effects). Thus, the *CUT1* coding region may be expressed under the control of the tapetum-specific promoters such as *TA29* (Mariani et al., 1990, 1992), *MS2* (Aarts et al., 1997), and *tap1* (Nacken et al., 1991).

Plant transformation vectors may also include RNA processing signals, for example, introns, which may be positioned upstream or downstream of the *CUT1* nucleic acid sequence in the transgene. In addition, the expression vectors may also include additional regulatory sequences from the 3'-untranslated region of plant genes, e.g., a 3' terminator region to increase mRNA stability of the mRNA, such as the *PI-II* terminator region of potato or the octopine or nopaline synthase 3' terminator regions.

Finally, as noted above, plant transformation vectors may also include dominant selectable marker genes to allow for the ready selection of transformants. Such genes include those encoding antibiotic resistance genes (e.g., resistance to hygromycin, kanamycin, bleomycin, G418, streptomycin or spectinomycin) and herbicide resistance genes (e.g., phosphinothricin acetyltransferase).

c. Arrangement of *CUT1* Nucleic Acids in Vector

As noted above, the particular arrangement of the *CUT1* nucleic acid in the transformation vector will be selected according to the expression of the nucleic acid desired.

Where enhanced VLCFA synthesis is desired, the *CUT1* nucleic acid may be operably linked to a constitutive high-level promoter such as the CaMV 35S promoter. Modification of VLCFA synthesis may also be achieved by introducing into a plant a transformation vector containing a variant form of the *CUT1* nucleic acid, for example a form which varies from the exact nucleotide sequence of the *CUT1* nucleic acid, but which encodes a protein that retains the functional characteristic of the *CUT1* protein, i.e., very long chain fatty acid elongation activity.

In contrast, a reduction of VLCFA synthesis may be obtained by introducing antisense constructs based on the *CUT1* nucleic acid sequence into plants. For antisense suppression, the *CUT1* nucleic acid is arranged in reverse orientation relative to the promoter sequence in the transformation vector. The introduced sequence need not be the full length *CUT1* nucleic acid, and need not be exactly homologous to the *CUT1* nucleic acid. Generally, however, where the introduced sequence is of shorter length, a higher degree of homology to the native *CUT1* sequence will be needed for effective antisense

suppression. Preferably, the introduced antisense sequence in the vector will be at least 30 nucleotides in length, and improved antisense suppression will typically be observed as the length of the antisense sequence increases. Preferably, the length of the antisense sequence in the vector will be greater than 100 nucleotides. Transcription of an antisense construct as described results in the production of RNA molecules that are the reverse complement of mRNA molecules transcribed from the endogenous *CUT1* gene in the plant cell. Although the exact mechanism by which antisense RNA molecules interfere with gene expression has not been elucidated, it is believed that antisense RNA molecules bind to the endogenous mRNA molecules and thereby inhibit translation of the endogenous mRNA.

Suppression of endogenous *CUT1* gene expression can also be achieved using ribozymes.

Ribozymes are synthetic RNA molecules that possess highly specific endoribonuclease activity. The production and use of ribozymes are disclosed in U.S. Patent No. 4,987,071 to Cech and U.S. Patent No. 5,543,508 to Haselhoff, which are hereby incorporated by reference. The inclusion of ribozyme sequences within antisense RNAs may be used to confer RNA cleaving activity on the antisense RNA, such that endogenous mRNA molecules that bind to the antisense RNA are cleaved, which in turn leads to an enhanced antisense inhibition of endogenous gene expression.

Constructs in which the *CUT1* nucleic acid (or variants thereon) are over-expressed may also be used to obtain co-suppression of the endogenous *CUT1* gene in the manner described in U.S. Patent No. 5,231,021 to Jorgensen. Such co-suppression (also termed sense suppression) does not require that the entire *CUT1* nucleic acid be introduced into the plant cells, nor does it require that the introduced sequence be exactly identical to the *CUT1* nucleic acid. However, as with antisense suppression, the suppressive efficiency will be enhanced as (1) the introduced sequence is lengthened and (2) the sequence similarity between the introduced sequence and the endogenous *CUT1* gene is increased. Example I below provides an illustration of co-suppression of the endogenous *CUT1* gene by transformation of plants with the *CUT1* cDNA.

d. Transformation and Regeneration Techniques

Transformation and regeneration of both monocotyledonous and dicotyledonous plant cells is now routine, and the selection of the most appropriate transformation technique will be determined by the practitioner. The choice of method will vary with the type of plant to be transformed; those skilled in the art will recognize the suitability of particular methods for given plant types. Suitable methods may include, but are not limited to: electroporation of plant protoplasts; liposome-mediated transformation; polyethylene mediated transformation; transformation using viruses; micro-injection of plant cells; micro-projectile bombardment of plant cells; vacuum infiltration; and *Agrobacterium tumefaciens* (AT) mediated transformation. Typical procedures for transforming and regenerating plants are described in the patent documents listed at the beginning of this section.

e. Selection of Transformed Plants

Following transformation and regeneration of plants with the transformation vector, transformed

plants are preferably selected using a dominant selectable marker incorporated into the transformation vector. Typically, such a marker will confer antibiotic resistance on the seedlings of transformed plants, and selection of transformants can be accomplished by exposing the seedlings to appropriate concentrations of the antibiotic. Example I provides an example of such an approach in which seedlings
 5 were selected using kanamycin.

After transformed plants are selected and grown to maturity, they can be assayed to determine whether VLCFA synthesis has been altered as a result of the introduced transgene. This can be done in several ways, including, as described in Example 1, microscopic examination of the epicuticular wax layer and chromatographic analysis. Lipids may also be extracted from plant material and analyzed by
 10 gas chromatography as described by Dooter (1990). In addition, antisense or sense suppression of the endogenous *CUT1* gene may be detected by analyzing mRNA expression on Northern blots.

IX. Production of Sequence Variants

As noted above, modification of VLCFA synthesis in plant cells can be achieved by
 15 transforming plants with *CUT1* nucleic acids, antisense constructs based on *CUT1* nucleic acid sequences or other variants on *CUT1* nucleic acid sequences. With the provision of the *CUT1* cDNA and genomic sequences herein, the creation of variants on these *CUT1* nucleic acid sequences by standard mutagenesis techniques is now enabled.

Variant DNA molecules include those created by standard DNA mutagenesis techniques, for
 20 example, M13 primer mutagenesis. Details of these techniques are provided in Sambrook et al. (1989), Ch. 15. By the use of such techniques, variants may be created which differ in minor ways from the disclosed *CUT1* nucleic acids. DNA molecules and nucleotide sequences that are derivatives of those specifically disclosed herein and which differ from those disclosed by the deletion, addition or substitution of nucleotides while still encoding a protein which possesses the functional characteristic of
 25 the *CUT1* protein (i.e., very long chain fatty acid elongation activity) are comprehended by this invention. DNA molecules and nucleotide sequences which are derived from the *CUT1* nucleic acids include DNA sequences which hybridize under moderately stringent conditions to the DNA sequences disclosed, or fragments thereof.

Hybridization conditions resulting in particular degrees of stringency will vary depending upon
 30 the nature of the hybridization method of choice and the composition and length of the hybridizing DNA used. Generally, the temperature of hybridization and the ionic strength (especially the Na⁺ concentration) of the hybridization buffer will determine the stringency of hybridization. Calculations regarding hybridization conditions required for attaining particular degrees of stringency are discussed by Sambrook et al. (1989), chapters 9 and 11, herein incorporated by reference. By way of illustration
 35 only, a hybridization experiment may be performed by hybridization of a *CUT1*-derived probe (for example, the *CUT1* cDNA sequence) to a target DNA molecule (for example, the *CUT1* homolog from *Zea Mays*) which has been electrophoresed in an agarose gel and transferred to a nitrocellulose membrane by Southern blotting (Southern, 1975), a technique well known in the art and described in

(Sambrook et al., 1989). Hybridization with a target probe labeled with [³²P]-dCTP is generally carried out in a solution of high ionic strength such as 6xSSC at a temperature that is 20-25°C below the melting temperature, T_m , described below. For such Southern hybridization experiments where the target DNA molecule on the Southern blot contains 10 ng of DNA or more, hybridization is typically carried out for 6-8 hours using 1-2 ng/ml radiolabeled probe (of specific activity equal to 10⁸ CPM/μg or greater). Following hybridization, the nitrocellulose filter is washed to remove background hybridization. The washing conditions should be as stringent as possible to remove background hybridization but to retain a specific hybridization signal. The term T_m represents the temperature above which, under the prevailing ionic conditions, the radiolabeled probe molecule will not hybridize to its target DNA molecule. The T_m of such a hybrid molecule may be estimated from the following equation (Bolton and McCarthy, 1962):

$$T_m = 81.5^\circ\text{C} - 16.6(\log_{10}[\text{Na}^+]) + 0.41(\%G+C) - 0.63(\% \text{ formamide}) - (600/l)$$

Where l = the length of the hybrid in base pairs.

This equation is valid for concentrations of Na^+ in the range of 0.01 M to 0.4 M, and it is less accurate for calculations of T_m in solutions of higher $[\text{Na}^+]$. The equation is also primarily valid for DNAs whose G+C content is in the range of 30% to 75%, and it applies to hybrids greater than 100 nucleotides in length (the behavior of oligonucleotide probes is described in detail in Ch. 11 of Sambrook et al., 1989).

Thus, by way of example, for a 150 base pair DNA probe derived from the first 150 base pairs of the open reading frame of the *CUT1* cDNA (with a hypothetical %GC = 45%), a calculation of hybridization conditions required to give particular stringencies may be made as follows:

For this example, it is assumed that the filter will be washed in 0.3 xSSC solution following hybridization, thereby $[\text{Na}^+] = 0.045\text{M}$, %GC = 45%, Formamide concentration = 0, $l = 150$ base pairs,

$$T_m = 81.5 - 16(\log_{10}[\text{Na}^+]) + (0.41 \times 45) - (600/150)$$

and so $T_m = 74.4^\circ\text{C}$.

The T_m of double-stranded DNA decreases by 1-1.5°C with every 1% decrease in homology (Bonner et al., 1973). Therefore, for this given example, washing the filter in 0.3 xSSC at 59.4-64.4°C will produce a stringency of hybridization equivalent to 90%. Alternatively, washing the hybridized filter in 0.3 xSSC at a temperature of 65.4-68.4°C will yield a hybridization stringency of 94%. The above example is given entirely by way of theoretical illustration. One skilled in the art will appreciate that other hybridization techniques may be utilized and that variations in experimental conditions will necessitate alternative calculations for stringency.

DNA sequences that encode a protein having VLCFA elongase activity and which hybridize to the disclosed *CUT1* nucleic acid sequences under hybridization conditions of at least 75%, more preferably at least 80%, 85% or 90% stringency, and most preferably at least 95% stringency are encompassed within the present invention.

The degeneracy of the genetic code further widens the scope of the present invention as it enables

major variations in the nucleotide sequence of a DNA molecule while maintaining the amino acid sequence of the encoded protein. For example, the fourth amino acid residue of the CUT1 protein is alanine. This is encoded in the *CUT1* ORF by the nucleotide codon triplet GCA. Because of the degeneracy of the genetic code, three other nucleotide codon triplets--GCT, GCC and GCG--also code for alanine. Thus, the nucleotide sequence of the *CUT1* ORF could be changed at this position to any of these three codons without affecting the amino acid composition of the encoded protein or the characteristics of the protein. Based upon the degeneracy of the genetic code, variant DNA molecules may be derived from the *CUT1* nucleic acid molecules disclosed herein using standard DNA mutagenesis techniques as described above, or by synthesis of DNA sequences. Thus, this invention also encompasses DNA sequences which encode the CUT1 protein but which vary from the *CUT1* nucleic acid sequences by virtue of the degeneracy of the genetic code.

One skilled in the art will recognize that DNA mutagenesis techniques may be used not only to produce variant DNA molecules, but will also facilitate the production of proteins which differ in certain structural aspects from the CUT1 protein, yet which proteins are clearly derivative of this protein and which maintain the essential characteristics of the CUT1 protein. Newly derived proteins may also be selected in order to obtain variations on the characteristic of the CUT1 protein, as will be more fully described below. Such derivatives include those with variations in amino acid sequence including minor deletions, additions and substitutions.

While the site for introducing an amino acid sequence variation is predetermined, the mutation *per se* need not be predetermined. For example, in order to optimize the performance of a mutation at a given site, random mutagenesis may be conducted at the target codon or region and the expressed protein variants screened for the optimal combination of desired activity. Techniques for making substitution mutations at predetermined sites in DNA having a known sequence as described above are well known.

Amino acid substitutions are typically of single residues; insertions usually will be on the order of about from 1 to 10 amino acid residues; and deletions will range about from 1 to 30 residues. Deletions or insertions preferably are made in adjacent pairs, i.e., a deletion of 2 residues or insertion of 2 residues. Substitutions, deletions, insertions or any combination thereof may be combined to arrive at a final construct. Obviously, the mutations that are made in the DNA encoding the protein must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure.

Substitutional variants are those in which at least one residue in the amino acid sequence has been removed and a different residue inserted in its place. Such substitutions generally are made in accordance with the following Table 1 when it is desired to finely modulate the characteristics of the protein. Table 1 shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative substitutions.

Table 1.

	Original Residue	Conservative Substitutions
5	Ala	ser
	Arg	lys
	Asn	gln; his
	Asp	glu
	Cys	ser
10	Gln	asn
	Glu	asp
	Gly	pro
	His	asn; gln
	Ile	leu, val
15	Leu	ile; val
	Lys	arg; gln; glu
	Met	leu; ile
	Phe	met; leu; tyr
	Ser	thr
20	Thr	ser
	Trp	tyr
	Tyr	trp; phe
	Val	ile; leu

Substantial changes in enzymatic function or other features are made by selecting substitutions that are less conservative than those in Table 1, i.e., selecting residues that differ more significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. The substitutions which in general are expected to produce the greatest changes in protein properties will be those in which (a) a hydrophilic residue, e.g., seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g., leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histadyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine.

The effects of these amino acid substitutions or deletions or additions may be assessed for derivatives of the CUT1 protein by analyzing the ability of the derivative proteins to catalyze the addition of C2 units to existing VLCFA units. These assays may conveniently be performed using the yeast-based systems for assaying fatty acid elongation described below.

X. Production of recombinant CUT1 protein using heterologous expression systems

Many different expression systems are available for expressing cloned nucleic acid molecules. Examples of prokaryotic and eukaryotic expression systems that are routinely used in laboratories are described in Chapters 16-17 of Sambrook et al. (1989), which are herein incorporated by reference. Such systems may be used to express CUT1 protein and derivatives at this protein at high levels to facilitate purification and functional analysis of the enzyme. Apart from permitting the activity of the enzyme to be

- determined (which is particularly useful to assess the activity of homologous and derivative proteins), heterologous expression facilitates other uses of the purified enzyme. For example the purified enzyme produced by recombinant means may be used to synthesize VLCFAs and other fatty acid metabolites in vitro, particularly radio- or fluorescent- labeled forms of VLCFAs and metabolites. These molecules may be used as tracers to determine the location in plant tissues and cells of VLCFAs and their metabolites. The purified recombinant enzyme may also be used as an immunogen to raise enzyme-specific antibodies. Such antibodies are useful as both research reagents (such as in the study of VLCFA regulation in plants) as well as diagnostically to determine expression levels of the enzyme in agricultural products, including pollen.
- By way of example only, high level expression of the CUT1 protein may be achieved by cloning and expressing the cDNA in yeast cells using the pYES2 yeast expression vector (Invitrogen, San Diego, CA). Secretion of the recombinant CUT1 from the yeast cells may be achieved by placing a yeast signal sequence adjacent to the CUT1 coding region. A number of yeast signal sequences have been characterized, including the signal sequence for yeast invertase. This sequence has been successfully used to direct the secretion of heterologous proteins from yeast cells, including such proteins as human interferon (Chang et al., 1986), human lactoferrin (Liang and Richardson, 1993) and prochymosin (Smith et al., 1985). Alternatively, the enzyme may be expressed at high level in standard prokaryotic expression systems, such as *E. coli*.

XI. Assays for VLCFA elongase activity

- To aid the biochemical characterization of the CUT1 protein, or variants of this protein, the very long chain fatty acid elongase activity of the proteins may be determined by expressing the cDNA molecule which encodes protein in question in yeast. For that purpose, the full-length coding region of the cDNA may be linked to the galactose inducible GAL1 promoter in the *Saccharomyces cerevisiae* expression vector, pYES2 (Invitrogen). The yeast expressing the subject protein may then be employed to determine the substrate specificity of the CUT1 protein by one of the following approaches.

a. In vitro assay for VLCFA elongase activity using cell-free yeast homogenate

- To determine the range of substrates recognized by the subject protein, acyl elongation activity is measured using substrates of varying carbon chain lengths and degrees of unsaturation. In each case, 15 μM of $[\text{1-}^{14}\text{C}]\text{acyl CoA}$ (C18, C20, C22, C24 in 0.005 % Triton X-100) is added to a standard assay mixture containing 80 mM Hepes-KOH, pH 7.2, 5 % glycerol, 1mM DTT, 0.5 mM NADPH, 1 mM ATP, 5 mM MgCl_2 , 1 mM malonyl-CoA, and an aliquot of cell free extract (50 μg protein) in a final volume of 50 μL . Incubation is carried out at 30°C for 1 h. The reaction is stopped with 100 μL of 4 N KOH in 80 % methanol and the lipids saponified for 1 h at 80°C. The mixture is then acidified by adding 100 μL of cold 6N HCL and extracted twice with 500 μL of cold hexane. The pooled hexane fractions are dried under N_2 , followed by transmethylolation for product analyses.

b. *In vivo* assay: Feeding of transformed yeast cells with radiolabelled acyl-tween substrates

A second approach for determining substrate specificity involves growth of yeast cells in the presence of various [$1\text{-}^{14}\text{C}$]acyl-Tween substrates (C18, C20, C22, C24; Terzaghi, 1986). Fatty acyl substrates provided in the growth medium as Tween-fatty acid esters are readily taken up from the medium and used by the cells. For each FAE protein, yeast cells are initially grown in the presence of several concentrations of a single acyl-Tween substrate for different lengths of time to determine the optimal substrate concentration and the duration of the feeding assays. Once these parameters are established, yeast cells expressing the subject protein and control cells containing empty pYES2 plasmid are grown in a defined medium in the presence of a single radiolabelled acyl-Tween substrate. At the end of the experiment, cells are pelleted, and then resuspended in 1 mL of 1 N methanolic-HCl (Supelco). Treatment with methanolic-HCl converts fatty acids to methyl esters (FAME). Radiolabelled FAMES are analyzed as described below, to characterize the products generated by elongation of each acyl-Tween substrate. A comparison of radiolabelled FAMES from CUT1 containing yeast with FAMES isolated from control cells allows the determination of the elongation specificity of the subject FAE protein.

c. Product analyses

The products of the elongation assays obtained in (a), or pelleted yeast cells from experiment (b) are transmethylated in a sealed tube using 1 N methanolic-HCl (Supelco) at 80°C for 1 h. Samples are then extracted twice with 500 μL of hexane after the addition of 1 mL of 0.9% NaCl, and the pooled extracts containing FAMES concentrated under N_2 . Radiolabelled FAMES are applied on KC_{18} reverse-phase TLC plates (Whatman), and separated in acetonitrile:tetrahydrofuran (85:15, v/v). Products of TLC separation are identified by co-chromatography with FAME standards, or by GC-MS. In addition, FAMES may be scraped from the TLC plates and their radioactivity determined by liquid scintillation counting.

EXAMPLES

The following examples serve to illustrate various applications of the present invention.

Example one: Modification of *A. thaliana* Wax Production By Transformation with the CUT1 cDNA

a. Construction of binary transformation vector

The *CUT1* cDNA was cleaved out of the vector λ ZipLox (with *KpnI*-*Bam*HI) and the resulting 1.85 kb fragment was directionally subcloned into the *KpnI*-*Bam*HI sites of pGEM7zf (Promega, Madison, WI). The resulting plasmid was then fully cleaved with *XhoI*, but only partially cleaved with *SsrI*, (since the *CUT1* cDNA has an internal *SsrI* site). The 1.9 kb product was isolated on an agarose gel and directionally subcloned into the *SalI* and *SsrI* sites of the vector pJD330 (Shaul and Galili 1992). This vector contains the 35S promoter of the cauliflower mosaic virus (CaMV) which provides constitutive expression in *Arabidopsis*. The subcloning results in the *CUT1* cDNA being inserted in a sense orientation with respect to the CaMV 35S promoter. The JD330-*CUT1* cDNA construct was ligated with pBIN19 and the resulting binary vector was designated p35S-*CUT1*. This binary vector was transformed into the *Agrobacterium tumefaciens* strain GV3101 (Koncz and Schell, 1986), and transformants were selected on LB medium containing 25 μ g/mL gentamycin and 50 μ g/mL kanamycin.

b. Transformation of *Arabidopsis* with the p35S-*CUT1* transgene

Arabidopsis thaliana (L.) Heynh. ecotype Columbia was transformed using a combination of *in planta* (Chang et al., 1994, Katavic et al., 1994) and vacuum infiltration methods (Bechtold et al., 1993). Plants were grown until the primary inflorescence shoots reached 1-2 cm in height, and then these bolts were cut off. The wound site was inoculated with 50 mL of an overnight *Agrobacterium* culture harbouring the p35S-*CUT1* plasmid. After 4-6 days a number of secondary inflorescences that appeared were cut off, and vacuum infiltration was performed on these plants using the conditions described by Bechtold et al. (1993). Screening for transformed seed was done as described previously (Katavic et al., 1994). Briefly, seed from infiltrated plants were plated out (approximately 1500 seeds/plate) on solid minimal salts nutrient medium supplemented with 50 μ g/mL kanamycin. Seedlings that showed resistance were visible after approximately 8 days, because they turned green and elongated. Plants that were derived from seed harvested from different pots were considered as independent lines. Designations of transformed plants were as follows: the infiltrated plant--T1; primary transformants--T2; etc., as outlined in Katavic et al. (1994). Plants were grown at 20°C under continuous fluorescent illumination (100 μ Em⁻²/s).

c. *CUT1*-suppressed plants have altered wax composition

Using the above transformation methods 46 kanamycin-resistant plants were obtained from seven different pots of *Arabidopsis*. Of the 46 plants obtained, 36 appeared waxless, having a glossy or *eceriferum* (*cer*) phenotype. At least one *cer* line was obtained from each pot implying that at least seven independent events had occurred in obtaining these lines. The surfaces of these *cer* plants were examined by a scanning electron (SE) microscope. SE micrographs clearly demonstrate that while wild-type plants were covered with the characteristic crystals of the epicuticular wax layer, transgenic *cer* plants were completely devoid of any wax crystals, implying that a severe *cer* phenotype has been created.

Plant tissue from the transgenic lines was analyzed for fatty acid composition. Plant tissue was immersed for 10 seconds in a 2:1 chloroform:methanol solution to remove surface waxes. Extracts were then evaporated to dryness under a stream of nitrogen. Waxes were dissolved in 100 μ l of *N,O*-bis(trimethylsilyl)trifluoroacetamide with 1% Trimethylchlorosilane (Pierce), and derivatized at 80 $^{\circ}$ C for 1 hour. Samples were analyzed in a Hewlett-Packard 5890 series II gas chromatograph equipped with a flame ionization detector, using either a DB-1 column or a DB-5 column.

GLC analyses were performed at the initial temperature of 150 $^{\circ}$ C, followed by a ramping of 4 $^{\circ}$ C/min to 320 $^{\circ}$ C, where it was held for 10 min. Peaks were identified by the comparison of retention times to reference standards, and mass spectrometry. Quantification was based on flame ionization detector peak areas, which were converted to mass units by comparison to the internal standard, 17:0-methylester, which was added to each sample prior to the extraction.

For wax load determinations only the principal surface lipids were measured, *n*-nonacosane (C29 alkane), 14- and 15-nonacosanol (C29 secondary alcohol), 15-nonacosanone (C29 ketone), C22-C30 aldehydes, C22-C30 primary alcohols and C16-C30 fatty acids (Hannoufa et al., 1993). The total area % of these peaks accounted for more than 90 % of the total area % of the sample.

The wax constituents that are found on the stems of *Arabidopsis* plants originate from two biosynthetic pathways (Figure 1). The decarbonylation pathway is the major pathway, which utilizes aldehydes to produce alkanes, secondary alcohols and ketones. In *Arabidopsis* (ecotype Columbia), the C29 species of the wax components produced by this pathway account for almost 90% of all the stem wax. The second pathway, the acyl-reduction pathway, produces primary alcohols, which account for approximately 5% of the total stem wax. Fatty acids and aldehydes, which are precursors for all the other wax components, are shared by both biosynthetic pathways and make up the remaining 5%.

Wax composition and quantity on the stems of wild-type and several transgenic lines were examined. Wild-type *Arabidopsis* stems contained on average 7106 (+/-) 1184 mg of wax/ g dry wt. In contrast, wax loads on the stems of all shiny *CUT1*-suppressed lines were severely reduced. For example, the wax load on the stems on the most severe line # 5 totals 483 (+/-) 83, only 6-7 % of the wild-type wax accumulation.

Analysis of wax composition of *CUT1*-suppressed plants revealed that the decarbonylation pathway is almost completely shut down. The C30 aldehyde, C29 alkane, C29 secondary alcohol and C29 ketone reach only 3.5 %, 2.2%, 1.4% and 2.2% of the levels found on wild-type plants, respectively. *CUT1*-suppression also has a major effect on the acyl-reduction pathway, causing a reduction in the levels of primary alcohols of over 50%. In addition, the relative abundance of different classes of alcohols is changed. C30 and C28 alcohols, the major alcohol species in wild type stems, have decreased by 90%, and C24 alcohol is the most abundant class in *CUT1* suppressed lines. The C24 species are also the most abundant classes of aldehydes and fatty acids in waxless transgenic plants. The described compositional changes were consistent in all 13 different *CUT1*-suppressed lines analyzed. These changes support the proposal that the role of the *CUT1* enzyme is elongation of the fatty acyl chain beyond 24 carbons.

Example two: Production of conditionally male sterile CUT1-suppressed plants

CUT1-suppressed *Arabidopsis* plants were produced as described in Example one and analyzed for male sterility. This analysis demonstrated that, in addition to stem and leaf wax synthesis, the *CUT1* gene product has an essential role in pollen development. Similar to *cer6-2* (Preuss et al., 1993) and *cer1* (Aarts et al., 1995) wax-deficient mutants of *Arabidopsis*, *CUT1*-suppressed plants are completely male sterile under normal growth conditions (30 to 40% relative humidity) although they produce normal amounts of pollen. However, when grown under high humidity (90 to 100%), pollen fertility is restored to the wild-type level, indicating that male sterility/fertility is conditional and environmentally controlled, just like in *cer6-2* and *cer1* mutants. For these two mutants, conditional male sterility is explained by alterations in the composition and content of the wax components of the tryphine layer covering the pollen grain. These long chain lipid molecules, produced in the tapetum layer of the anther, (Preuss et al., 1993) are needed in the tryphine for proper pollen-pistil signalling and pollen germination. Thus, in their absence, sterility occurs. Conditional male sterility is a valuable trait for plant breeders; being able to selectively inhibit self-fertilization of plants facilitates the production of hybrid plants. Accordingly, the *CUT1* cDNA and derivatives thereof may be useful in producing conditionally male sterile plants useful in breeding programs.

Taken together, the results of Examples one and two confirm that *CUT1* encodes a condensing enzyme that is involved in VLCFA biosynthesis of waxes which accumulate in the plant epidermis, as well as waxes required for the development of functional pollen grains. In addition the results show that transformation of plants using the *CUT1* cDNA is useful to produce plants having modified VLCFA compositions, as well as plants that exhibit conditional male sterility.

Example three: Use of CUT1 gene promoter sequence

The promoter of the *CUT1* gene confers epidermis-specific expression. Accordingly, this promoter sequence may be used to produce transgene constructs that are specifically expressed in epidermal cells. Effective epidermis-specific expression may be achieved with less than the entire 1951 bases of sequence upstream of the *CUT1* ORF shown in Seq. I.D. No. 12. Thus, by way of example, epidermis-specific expression may be obtained by employing the 1209 base pair promoter fragment. One of skill in the art will recognize that still smaller regions of the sequence upstream of the *CUT1* ORF may be used to obtain epidermis-specific expression, such as a 50 base pair or 100 base pair region of the disclosed promoter sequence.

The determination of whether a particular sub-region of the disclosed sequence operates to confer effective epidermis-specific expression in a particular system (taking into account the plant species into which the construct is being introduced, the level of expression required, etc.) will be performed using known methods, such as operably linking the promoter sub-region to a marker gene (e.g. GUS), introducing such constructs into plants and then determining the level of expression of the marker gene in epidermis and other

plant tissues.

The present invention therefore facilitates the production, by standard molecular biology techniques, of nucleic acid molecules comprising this promoter sequence operably linked to a nucleic acid sequence, such as an open reading frame. Suitable open reading frames include open reading frames encoding any protein

5 for which epidermis-specific expression is desired.

Having illustrated and described the principles of isolating *CUT1* nucleic acids, the *CUT1* protein and modes of use of these biological molecules, it should be apparent to one skilled in the art that the invention can be modified in arrangement and detail without departing from such principles. We claim all

10 modifications coming within the spirit and scope of the claims presented herein.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: The University of British Columbia
- 5 (ii) TITLE OF INVENTION: Nucleic Acids Encoding Plant Enzyme Involved In Very Long Chain Fatty Acid Synthesis
- (iii) NUMBER OF SEQUENCES: 12
- (iv) CORRESPONDENCE ADDRESS:
- 10 (A) ADDRESSEE: Sim & McBurney
- (B) STREET: 6th Floor, 330 University Avenue
- (C) CITY: Toronto
- (D) PROVINCE: Ontario
- (E) COUNTRY: Canada
- (F) POSTAL CODE: M5G 1R7
- 15 (v) COMPUTER READABLE FORM:
- (A) MEDIUM TYPE: Disk, 3.5-inch
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: Windows 95
- (D) SOFTWARE: ASCII
- 20 (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE: 14 April 1998
- (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
- 25 (A) APPLICATION NUMBER: 60/043,831
- (B) FILING DATE: April 14, 1997
- (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE: April 10, 1998
- 30 (viii) PATENT AGENT INFORMATION
- (A) NAME: RAE, Patricia A.
- (B) REFERENCE/DOCKET NUMBER: 3055-18/PAR
- (ix) TELECOMMUNICATION INFORMATION:
- (A) TELEPHONE: (416) 595-1155
- 35 (B) TELEFAX: (416) 595-1163
- (2) INFORMATION FOR SEQ ID NO: 1:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 3712
- 40 (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:
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- 50 ATCGTAACTA CTGTTTGATT TTCCTCATG CTTGCAGTTT TCATGGATAT 200
- CTCAAGATTG GTCTTCTTGC ACTTCCCAAG TCAAACATAA AGTAACTACT 250

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	Pro Thr Met Asp Ala Ala Arg Ser Glu Ala Gln Met Val Ile	
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15	CCT AAA GAC GTC GAC ATC CTT ATC GTC AAC TGC TCT CTT TTC	2536
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	Lys Tyr Lys Leu Ser His Leu Val Arg Thr His Arg Gly Ala	
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	Asp Asp Lys Ser Phe Tyr Cys Val Tyr Glu Gln Glu Asp Lys	
	310 315 320	
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	Pro Leu Val Leu Pro Ala Ser Glu Gln	Leu Leu Phe Leu Thr		
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	Ser Leu Ile Gly Arg Lys Ile Phe Asn	Pro Lys Trp Lys Pro		
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	(A) LENGTH: 1807	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: double	
	(D) TOPOLOGY: linear	
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	Ala Val Glu Leu Leu Arg Met Gly Pro Glu Glu Ile Leu Asn	
	45 50 55	
	GTT TGG AAT TCA CTC CAG TTT GAC CTA GTT CAG GTT CTA TGT	207
	Val Trp Asn Ser Leu Gln Phe Asp Leu Val Gln Val Leu Cys	
	60 65	
40	TCT TCC TTC TTT GTC ATC TTC ATC TCC ACT GTT TAC TTC ATG	249
	Ser Ser Phe Phe Val Ile Phe Ile Ser Thr Val Tyr Phe Met	
	70 75 80	
45	TCC AAG CCA CGC ACC ATC TAC CTC GTT GAC TAT TCT TGT TAC	291
	Ser Lys Pro Arg Thr Ile Tyr Leu Val Asp Tyr Ser Cys Tyr	
	85 90 95	
50	AAG CCA CCT GTC ACG TGT CGT GTC CCC TTC GCA ACT TTC ATG	333
	Lys Pro Pro Val Thr Cys Arg Val Pro Phe Ala Thr Phe Met	
	100 105 110	
	GAA CAC TCT CGT TTG ATC CTC AAG GAC AAG CCT AAG AGC GTC	375
	Glu His Ser Arg Leu Ile Leu Lys Asp Lys Pro Lys Ser Val	
	115 120 125	

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	GAG TTC CAA ATG AGA ATC CTT GAA CGT TCT GGC CTC GGT GAG	417
	Glu Phe Gln Met Arg Ile Leu Glu Arg Ser Gly Leu Gly Glu	
	130 135	
5	GAG ACT TGT CTC CCT CCG GCT ATT CAT TAT ATT CCT CCC ACA	459
	Glu Thr Cys Leu Pro Pro Ala Ile His Tyr Ile Pro Pro Thr	
	140 145 150	
10	CCA ACC ATG GAC GCG GCT AGA AGC GAG GCT CAG ATG GTT ATC	501
	Pro Thr Met Asp Ala Ala Arg Ser Glu Ala Gln Met Val Ile	
	155 160 165	
	TTC GAG GCC ATG GAC GAT CTT TTC AAG AAA ACC GGT CTT AAA	543
15	Phe Glu Ala Met Asp Asp Leu Phe Lys Lys Thr Gly Leu Lys	
	170 175 180	
	CCT AAA GAC GTC GAC ATC CTT ATC GTC AAC TGC TCT CTT TTC	585
20	Pro Lys Asp Val Asp Ile Leu Ile Val Asn Cys Ser Leu Phe	
	185 190 195	
	TCT CCC ACA CCA TCG CTC TCA GCT ATG GTC ATC AAC AAA TAT	627
	Ser Pro Thr Pro Ser Leu Ser Ala Met Val Ile Asn Lys Tyr	
	200 205	
25	AAG CTT AGG AGT AAT ATC AAG AGC TTC AAT CTT TCG GGG ATG	669
	Lys Leu Arg Ser Asn Ile Lys Ser Phe Asn Leu Ser Gly Met	
	210 215 220	
30	GGC TGC AGC GCG GCG CTG ATC TCA GTT GAT CTA GCC CGC GAC	711
	Gly Cys Ser Ala Gly Leu Ile Ser Val Asp Leu Ala Arg Asp	
	225 230 235	
	TTG CTC CAA GTT CAT CCC AAT TCA AAT GCA ATC ATC GTC AGC	753
35	Leu Leu Gln Val His Pro Asn Ser Asn Ala Ile Ile Val Ser	
	240 245 250	
	ACG GAG ATC ATA ACG CCT AAT TAC TAT CAA GGC AAC GAG AGA	795
40	Thr Glu Ile Ile Thr Pro Asn Tyr Tyr Gln Gly Asn Glu Arg	
	255 260 265	
	GCC ATG TTG TTA CCC AAT TGT CTC TTC CGC ATG GGT GCG GCA	837
	Ala Met Leu Leu Pro Asn Cys Leu Phe Arg Met Gly Ala Ala	
	270 275	
45	GCC ATA CAC ATG TCA AAC CGC CGG TCT GAC CGG TGG CGA GCC	879
	Ala Ile His Met Ser Asn Arg Arg Ser Asp Arg Trp Arg Ala	
	280 285 290	
50	AAA TAC AAG CTT TCC CAC CTC GTC CGG ACA CAC CGT GGC GCT	921
	Lys Tyr Lys Leu Ser His Leu Val Arg Thr His Arg Gly Ala	
	295 300 305	
	GAC GAC AAG TCT TTC TAC TGT GTC TAC GAA CAG GAA GAC AAA	963

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	Asp Asp Lys Ser Phe Tyr Cys Val Tyr Glu Gln Glu Asp Lys	
	310 315 320	
5	GAA GGA CAC GTT GGC ATC AAC TTG TCC AAA GAT CTC ATG GCC Glu Gly His Val Gly Ile Asn Leu Ser Lys Asp Leu Met Ala	1005 335
	325 330	
10	ATC GCC GGT GAA GCC CTC AAG GCA AAC ATC ACC ACA ATA GGT Ile Ala Gly Glu Ala Leu Lys Ala Asn Ile Thr Thr Ile Gly	1047 345
	340	
15	CCT TTG GTC CTA CCG GCG TCA GAA CAA CTT CTC TTC CTC ACG Pro Leu Val Leu Pro Ala Ser Glu Gln Leu Leu Phe Leu Thr	1089 360
	350 355	
	TCC CTA ATC GGA CGT AAA ATC TTC AAC CCG AAA TGG AAA CCA Ser Leu Ile Gly Arg Lys Ile Phe Asn Pro Lys Trp Lys Pro	1131 375
	365 370	
20	TAC ATA CCG GAT TTC AAG CTG GCC TTC GAA CAC TTT TGC ATT Tyr Ile Pro Asp Phe Lys Leu Ala Phe Glu His Phe Cys Ile	1173 390
	380 385	
25	CAC GCA GGA GGC AGA GCG GTG ATC GAC GAG CTC CAA AAG AAT His Ala Gly Gly Arg Ala Val Ile Asp Glu Leu Gln Lys Asn	1215 405
	395 400	
30	CTA CAA CTA TCA GGA GAA CAC GTT GAG GCC TCA AGA ATG ACA Leu Gln Leu Ser Gly Glu His Val Glu Ala Ser Arg Met Thr	1257 415
	410	
35	CTA CAT CGT TTT GGT AAC ACG TCA TCT TCA TCG TTA TGG TAC Leu His Arg Phe Gly Asn Thr Ser Ser Ser Leu Trp Tyr	1299 430
	420 425	
	GAG CTT AGC TAC ATC GAG TCT AAA GGG AGA ATG AGG AGA GGC Glu Leu Ser Tyr Ile Glu Ser Lys Gly Arg Met Arg Arg Gly	1341 445
	435 440	
40	GAT CGC GTT TGG CAA ATC GCG TTT GGG AGT GGT TTC AAG TGT Asp Arg Val Trp Gln Ile Ala Phe Gly Ser Gly Phe Lys Cys	1383 460
	450 455	
45	AAC TCT GCC GTG TGG AAA TGT AAC CGT ACG ATT AAG ACA CCT Asn Ser Ala Val Trp Lys Cys Asn Arg Thr Ile Lys Thr Pro	1425 475
	465 470	
50	AAG GAC GGA CCA TGG TCC GAT TGT ATC GAC CGT TAC CCT GTC Lys Asp Gly Pro Trp Ser Asp Cys Ile Asp Arg Tyr Pro Val	1467 485
	480	
	TTT ATT CCC GAA GTT GTC AAA CTC Phe Ile Pro Glu Val Val Lys Leu	1491

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(2) INFORMATION FOR SEQ ID NO: 4

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 497

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

```

Met Pro Gln Ala Pro Met Pro Glu Phe Ser Ser Ser Val Lys Leu Lys
10
1      5      10      15
Tyr Val Lys Leu Gly Tyr Gln Tyr Leu Val Asn His Phe Leu Ser Phe
20
Leu Leu Ile Pro Ile Met Ala Ile Val Ala Val Glu Leu Leu Arg Met
15
35      40      45
Gly Pro Glu Glu Ile Leu Asn Val Trp Asn Ser Leu Gln Phe Asp Leu
50      55      60
Val Gln Val Leu Cys Ser Ser Phe Phe Val Ile Phe Ile Ser Thr Val
20
65      70      75      80
Tyr Phe Met Ser Lys Pro Arg Thr Ile Tyr Leu Val Asp Tyr Ser Cys
85      90      95
Tyr Lys Pro Pro Val Thr Cys Arg Val Pro Phe Ala Thr Phe Met Glu
25
100      105      110
His Ser Arg Leu Ile Leu Lys Asp Lys Pro Lys Ser Val Glu Phe Gln
115      120      125
Met Arg Ile Leu Glu Arg Ser Gly Leu Gly Glu Glu Thr Cys Leu Pro
30
130      135      140
Pro Ala Ile His Tyr Ile Pro Pro Thr Pro Thr Met Asp Ala Ala Arg
145      150      155      160
Ser Glu Ala Gln Met Val Ile Phe Glu Ala Met Asp Asp Leu Phe Lys
35
165      170      175
Lys Thr Gly Leu Lys Pro Lys Asp Val Asp Ile Leu Ile Val Asn Cys
180      185      190
Ser Leu Phe Ser Pro Thr Pro Ser Leu Ser Ala Met Val Ile Asn Lys
40
195      200      205
Tyr Lys Leu Arg Ser Asn Ile Lys Ser Phe Asn Leu Ser Gly Met Gly
45
210      215      220
Cys Ser Ala Gly Leu Ile Ser Val Asp Leu Ala Arg Asp Leu Leu Gln
225      230      235      240
Val His Pro Asn Ser Asn Ala Ile Ile Val Ser Thr Glu Ile Ile Thr
50
245      250      255
Pro Asn Tyr Tyr Gln Gly Asn Glu Arg Ala Met Leu Leu Pro Asn Cys
260      265      270

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Leu Phe Arg Met Gly Ala Ala Ala Ile His Met Ser Asn Arg Arg Ser
 275 280 285
 5 Asp Arg Trp Arg Ala Lys Tyr Lys Leu Ser His Leu Val Arg Thr His
 290 295 300
 Arg Gly Ala Asp Asp Lys Ser Phe Tyr Cys Val Tyr Glu Gln Glu Asp
 305 310 315 320
 10 Lys Glu Gly His Val Gly Ile Asn Leu Ser Lys Asp Leu Met Ala Ile
 325 330 335
 Ala Gly Glu Ala Leu Lys Ala Asn Ile Thr Thr Ile Gly Pro Leu Val
 340 345 350
 15 Leu Pro Ala Ser Glu Gln Leu Leu Phe Leu Thr Ser Leu Ile Gly Arg
 355 360 365
 20 Lys Ile Phe Asn Pro Lys Trp Lys Pro Tyr Ile Pro Asp Phe Lys Leu
 370 375 380
 Ala Phe Glu His Phe Cys Ile His Ala Gly Gly Arg Ala Val Ile Asp
 385 390 395 400
 25 Glu Leu Gln Lys Asn Leu Gln Leu Ser Gly Glu His Val Glu Ala Ser
 405 410 415
 Arg Met Thr Leu His Arg Phe Gly Asn Thr Ser Ser Ser Ser Leu Trp
 420 425 430
 30 Tyr Glu Leu Ser Tyr Ile Glu Ser Lys Gly Arg Met Arg Arg Gly Asp
 435 440 445
 35 Arg Val Trp Gln Ile Ala Phe Gly Ser Gly Phe Lys Cys Asn Ser Ala
 450 455 460
 Val Trp Lys Cys Asn Arg Thr Ile Lys Thr Pro Lys Asp Gly Pro Trp
 465 470 475 480
 40 Ser Asp Cys Ile Asp Arg Tyr Pro Val Phe Ile Pro Glu Val Val Lys
 485 490 495
 Leu
 497

45

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

GTGCTTTATA TATGTTTG 18

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- (2) INFORMATION FOR SEQ ID NO: 6:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 18
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:
- 10 CGTCGGAGAG TTTTAATG 18
- (2) INFORMATION FOR SEQ ID NO: 7:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 18
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:
- 20 CTCGATATC GGTGTTG 18
- (2) INFORMATION FOR SEQ ID NO: 8:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 24
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:
- 30 .AAATACCCTA ATCACATTT GTAA 24
- (2) INFORMATION FOR SEQ ID NO: 9:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 24
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:
- 40 TTTAACAGA GAGAAATATT CTTA 24
- (2) INFORMATION FOR SEQ ID NO: 10:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 24
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:
- 50 ATGCCTCAGG CACCGATGCC AGAG 24
- (2) INFORMATION FOR SEQ ID NO: 11:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 24

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(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

5 CAGCACGAGA AACTAAAAAA TACC 24

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

10 (A) LENGTH: 1951
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

15 TAGTGCTTTA TATATGTTTG ATACTTCTGT TTGGCAATAT CAATCATAGT 50
 AGAAAAGATA TGGACTTCAT TTGAGGTTTT TGGTGGATTG TGCTATATG 100
 20 TGAATCATG GGATCTCAAG ATTTGTCTGC ATTCAGTTTC CAAGTCAAAC 150
 ATCGTAAC TA CTGTTGATT TTCCCTCATG CTTGCAGTTT TCATGGATAT 200
 25 CTCAGATTG GTCTTCTTGC ACTTTCCAAG TCAAACATAA AGTAAC TACT 250
 GATTGATATT CCCTCGTGTA TTACCCCTCTT TCAAATGACA CAATTGGGCC 300
 CAAGTAGAGG AATTCATAG TGAATTCAAA AGATTAACTG TATCCACCG 350
 30 TCGTATTTTG ATAACATTTA GTTATTCCTT TCTTTTTTTT TCTCTGCAA 400
 CAGTTTTTTT TTAATACATT TAGTGTGGT TTGGTCAAT GAAATATTAT 450
 ATGTTACTTC TTTTTTTGGA AATAAATTAT TCATCTTTC TACTATAAAA 500
 35 GGAATTGTTT ATGCTTTTTT GATACAATAG TATACCATT TCAAAGATAC 550
 CATAGACCAG TTATTACATG AATCGCCAAA ACAACACTAA AATCAGAAAA 600
 40 TCAATATATT TTGGTATAGT CTCACACATA CAATCATAAA ACCTCTGTGA 650
 AATTAAAAAT CTATATTGTA CATTTCAAAG TTAAACAACA TAGTCTTAAA 700
 TAATTACCTA AATTTTAAAGT CAAATGTGAA TTATATTTTA CTCCTCGATA 750
 45 TCGGTTGTTG ACGATTAAAC ATGCAAAAAA GAAACATTAA TTGCGAATGT 800
 AAATAACAAA ACATGTAAC TTTGTAGATA TACATGTATC GACATTTAAA 850
 50 CCCGAATATA TATGTATACC TATAATTCTC CTGATTTTCA CGCTACCTGC 900
 CACGTACATG GGTGATAGGT CCAAACCTCAC AAGTAAAAGT TTACGTACAG 950
 TGAATTCGTC TTTTGGGTA TAAACGTACA TTTAATTAC ACGTAAGAAA 1000

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	GGATTACCAA	TCTTTTCATT	TATGGTACCA	GACAGAGTTA	AGGCAAACAA	1050
	GAGAAACATA	TAGAGTTTTG	ATATGTTTTC	TTGGATAAAT	ATTAAATTGA	1100
5	TCGAATATTT	AGGGATGGAC	ACAAGGTAAT	ATATGCCTTT	TAAGGTATAT	1150
	GTGCTATATG	AATCGTTTCG	CATGGGTACT	AAAATTATTT	GTCCTTACTT	1200
10	TATATAACA	AATTCCAACA	AAATCAAGTT	TTTGCTAAAA	CTAGTTTATT	1250
	TGCGGGTTAT	TTAATTACCT	ATCATATTAC	TTGTAATATC	ATTCGTATGT	1300
	TAACGGGTAA	ACCAAACCAA	ACCGGATATT	GAACATTAA	AAATCTTGTA	1350
15	AAATTGACAC	AAACTAATGA	ATATCTAAAT	TATGTTACTG	CTATGATAAC	1400
	GACCATTTTT	GTTTTTGAGA	ACCATAATAT	AAATTACAGG	TACGTGACAA	1450
20	GTACTAAGTA	TTTATATCCA	CCTTTAGTCA	CAGTACCAAT	ATTGCGCCTA	1500
	CCGGGCAACG	TGAACGTGAT	CATCAAATCA	AAGTAGTTAC	CAAACGCTTT	1550
	GATCTCGATA	AACTAAAAG	CTGACACGTC	TTGCTGTTTC	TTAATTTATT	1600
25	TCTCTTACAA	CGACAATTTT	GAGAAATATG	AAATTTTAT	ATCGAAAGGG	1650
	AACAGTCCTT	ATCATTTGCT	CCCATCACIT	GCITTTGTCT	AGTTACAAC	1700
30	GGAAATCGAA	GAGAAGTATT	ACAAAAACAT	TTTTCTCGTC	ATTTATAAAA	1750
	AAATGACAAA	AAATTAAATA	GAGAGCAAAG	CAAGAGCGTT	GGGTGACGTT	1800
	GGTCTCTTCA	TTAACTCCTC	TCACTACCC	CTTCCTCTGT	TGCGCTTTAT	1850
35	ATCCTTCACC	TTCCCTCTCT	CATCTTCATT	AACTCATCTT	CAAAAAATACC	1900
	CTAATCACAT	TTTGTAACAA	TAATACAATT	ATACATTAAA	ACTCTCCGAC	1950
40	G					1951

45

ANNEX

Claims

We claim:

1. An isolated nucleic acid molecule that encodes a protein having very long chain fatty acid elongase activity, wherein the nucleic acid molecule is selected from the group consisting of:
- (a) nucleic acids comprising at least 15 consecutive nucleotides of the sequence set forth in Seq. I.D. No. 3;
 - (b) nucleic acids possessing at least 70% sequence identity with the sequence set forth in Seq. I.D. No. 3, and
 - (c) nucleic acids that hybridize under conditions of at least 70% stringency with the sequence set forth in Seq. I.D. No. 3.
2. An isolated nucleic acid molecule according to claim 1 wherein the nucleic acid molecule comprises the sequence set forth in Seq. I.D. No. 3.
3. An isolated nucleic acid molecule according to claim 1 wherein the nucleic acid molecule possess at least 80% sequence identity with the sequence set forth in Seq. I.D. No. 3.
4. An isolated nucleic acid molecule according to claim 1 wherein the nucleic acid molecule hybridizes under conditions of at least 80% stringency with the sequence set forth in Seq. I.D. No. 3.
5. A purified protein encoded by a nucleic acid molecule according to claim 1, wherein the purified protein has an amino acid sequence having greater than 50 percent sequence identity with the amino acid sequence shown in Seq. I.D. No. 4.
6. A recombinant vector comprising a nucleic acid molecule according to claim 1

7. A recombinant vector according to claim 6 wherein the nucleic acid molecule is in reverse orientation relative to an adjacent promoter sequence of the vector.

5 8. A transgenic plant comprising a recombinant vector according to claim 6.

9. A transgenic plant comprising a recombinant vector according to claim 7.

10

10. A transgenic plant comprising a recombinant expression cassette comprising a promoter sequence operably linked to a nucleic acid sequence selected from the group consisting of:

15 (a) nucleic acids comprising at least 15 consecutive nucleotides of the sequence set forth in Seq. I.D. No. 3;

(b) nucleic acids possessing at least 70% sequence identity with the sequence set forth in Seq. I.D. No. 3; and

(c) nucleic acids that hybridize under conditions of at least 70% stringency with the sequence set forth in Seq. I.D. No. 3.

20

11. A transgenic plant according to claim 10 wherein the nucleic acid sequence comprises at least 30 consecutive nucleotides of the sequence set forth in Seq. I.D. No. 1.

25

12. A transgenic plant according to claim 10 wherein the nucleic acid sequence possess at least 80% sequence identity with the sequence set forth in Seq. I.D. No. 3.

30

13. A transgenic plant according to claim 10 wherein the nucleic acid sequence hybridizes under conditions of at least 80% stringency with the sequence set forth in Seq. I.D. No. 3.

14. A transgenic plant according to claim 10, wherein the plant has a modified phenotype compared to a non-transgenic plant of the same species.

15 15. A transgenic plant according to claim 14 wherein the modified phenotype is a modified very long chain fatty acid composition.

16. A transgenic plant according to claim 15 wherein the modified phenotype is a modified epicuticular wax layer.

10 17. A transgenic plant according to claim 14 wherein the modified phenotype is modified seed oil composition.

18. A transgenic plant according to claim 14 wherein the modified phenotype is conditional male sterility.

15

19. A method of producing a plant with a modified very long chain fatty acid composition relative to a non-transgenic plant of the same species, comprising introducing into the plant a recombinant vector according to claim 6.

20 20. A transgenic plant produced by the method of claim 19.

21. A transgenic plant produced by sexual or asexual propagation of a plant according to claim 20 or the progeny of said plant.

25 22. An isolated nucleic acid molecule having a nucleotide sequence according to Seq. I.D. No. 3.

23. An isolated nucleotide that encodes a protein having an amino acid sequence as shown in Seq. I.D. No. 4.

30

24. A method of isolating a nucleic acid molecule encoding a plant very long chain fatty acid elongation enzyme, the method comprising hybridizing a nucleic acid preparation with a DNA molecule comprising at least 15 consecutive nucleotides of the sequence set forth in Seq. I.D. No. 3

5

25. An isolated nucleic acid molecule isolated according to the method of claim 24.

26. A recombinant nucleic acid molecule comprising a promoter
10 sequence operably linked to a nucleic acid sequence, wherein the promoter sequence comprises a CUT1 promoter.

27. A recombinant nucleic acid molecule according to claim 26 wherein the promoter sequence comprises at least 50 consecutive nucleotides of the sequence
15 shown in Seq. I.D. No. 12.

28. A purified peptide having an amino acid sequence that is at least 70% identical to the sequence set forth in Seq. I.D. No. 4.

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Arabidopsis Wax Biosynthesis

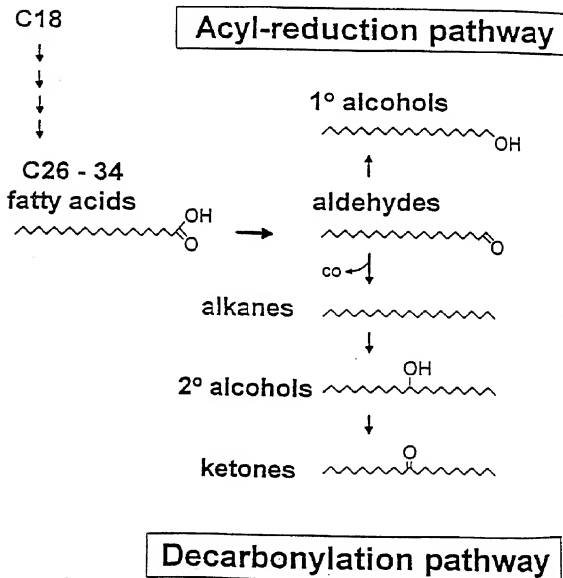


FIG. 1